TheGMS



The Gazette of Medical Sciences

https://www.thegms.co

ISSN 2692-4374

DOI https://www.doi.org/10.46766/thegms

Neurology | Review

COVID-19: Energy, Protein **Folding & Prion Disease**

K. E. Thorp¹, James A. Thorp^{2*}, Elise M. Thorp³, Margery M. Thorp⁴, Paul R. Walker⁵

¹MD. Department of Radiology, Sparrow Health System, Lansing, MI.

²MD. Department of Obstetrics and Gynecology, Division of Maternal Fetal Medicine, Sisters of St. Mary's Health System, St. Louis, MO.

³BS, FNTP. Williamston, MI.

⁴JD MACP. Law Firm of Margery M. Thorp, PLLC, Gulf Breeze, FL.

⁵BSME, MSEE

Submitted: 31 August 2022 Approved: 12 September 2022

Published: 13 September 2022

Address for correspondence:

James A. Thorp, Department of Obstetrics and Gynecology, Division of Maternal Fetal Medicine, Sisters of St. Mary's Health System, St. Louis, MO

How to cite this article: Thorp KE, Thorp JA, Thorp EM, Thorp MM, Walker PR. COVID-19: Energy, Protein Folding & Prion Disease. G Med Sci. 2022; 3(1): 0179-0206. https://www.doi.org/10.46766/thegms.neuro.22083101

Copyright: © 2022 K. E. Thorp, James A. Thorp, Elise M. Thorp, Margery M. Thorp, Paul R. Walker. This is an Open Access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Introduction

The recent recognition of intravascular amyloid formation with deposition of insoluble microthrombi throughout the circulatory system in primary COVID-19 infection or following administration of mRNA vaccines is a pivotal discovery that alters conventional notions about the nature of the underlying pathologic process at play in SARS-CoV-2 infection.

Since early in the pandemic researchers have ascribed the cascade of pathology to intravascular coagulation induced by the so-called cytokine storm and related immune system dysfunction. Alongside the web of deterioration, studies chronicle a range of autoantibodies, a dozen or more in some individuals, directed against proteins like cytokines, chemokines, cell surface proteins as well as RNA and DNA. Antibodies to platelet factor 4, for example, are believed to play a key role in the propagation of clot within the vascular system. While such reports must be regarded as factual, they are merely descriptions that do not, in and of themselves, explain the progressively widening arc of intravascular pathology. The presence of amyloid, however, compels one to fundamentally reconsider the nature of the problem.

Amyloid deposition can occur in any organ of the body and is associated with an increasing number of pathologic states including diseases like Alzheimer's, Parkinson's as well as type II diabetes. Amyloid is composed of singlestranded protein fibrils held together by hydrogen bonds to form the characteristic birefringent β-sheet structure. At least 30-40 proteins have been identified as precursors to the amyloid state but in any given patient or disease condition the amyloid usually derives from but a single type. Protein misfolding is widely regarded to be the primary cause of amyloid aggregation and deposition.

Protein misfolding is now regarded as a leading cause of chronic disease and has been associated with the neurodegenerative diseases, diabetes, cystic fibrosis, sickle cell anemia, as well as various cancers. All functional activities in the body - movement, nerve transmission, molecular transport, secretion, cellular division, enzyme activities and more - are effected by conformational changes in folded proteins. Even the appearance of autoantibodies in the blood during COVID-19 infections can be explained on the basis of protein misfolding.

But the folding of proteins into their three-dimensional functional structure is an energy-dependent process and disruption of intracellular energy availability necessarily



impacts folding. Conversely, accumulation of amyloid as well as the characteristic intracellular aggregates in disease states such as Alzheimer's or Parkinson's reflect impairment of lysosomal function and intracellular digestive pathways, i.e., autophagy, which are also dependent upon sustained energy flow.

In recent decades there has been increasing focus on a rare class of disorder known as the prion diseases, progressive and invariably fatal neurodegenerative conditions. The pathologic chain of events in the central nervous system (CNS) are a result of protein misfolding caused by propagation a pathogenic misfolded form of a naturally occurring protein inside the body.

While little is known of how prion particles propagate through nerve cells or induce misfolding of native proteins, the prion concept itself is extremely useful and would seem to account for how a wide range of protein misfolding conditions ultimately develop and progress. Prion-like mechanisms have increasingly been invoked to explain the origins of Alzheimer's and Parkinson's diseases as well as other conditions including amyloid deposition. Not surprisingly, some have suggested that the SARS-CoV-2 spike protein produces its pathological sequelae through prion-like mechanisms.

In this article we focus on the disturbed energy milieu associated with COVID-19 infection and show how all manifestations of the syndrome, from the earliest prodromal constitutional symptoms such as fever, malaise and lethargy, to more advanced morbid alterations such as intravascular thrombosis and organ dysfunction, can be explained on the basis of a mounting energy debt. Moreover, functional alterations such as autoantibodies, clotting disturbances, and organ failure ultimately trace back to protein misfolding and impairment of conformational change in affected proteins.

Cardiovascular Energy Dynamics

One of the most far-reaching and impactful conceptual turn of events in 20th century medicine occurred in the 1980s without most medical scientists or physicians even recognizing that the ground beneath their feet had begun to crumble. A decisive reshaping of ideas was underway that, ultimately, would render the molecular and cellular framework obsolete. This profound intellectual transformation constitutes what science historian Thomas Kuhn, in his insightful work *The Structure of Scientific*

Revolutions (1962), described as a paradigm shift [1].

For much of the 20th century scientists conceived the heart to function in the manner of a mechanical pump, with blood propelled forward through the arteries during the systolic (contraction) phase of the cardiac cycle. The diastolic phase of the cycle, conversely, was believed to represent a period of passive relaxation. This notion was originally advanced by William Harvey in 1628 in his seminal work *On the Motions of the Heart* in which he describes his discovery of the circulation of the blood [2]. Harvey's model was uncritically adopted by English physiologist Ernest Starling in the early 20th century and thereafter became accepted as fact. The problem with Starling's so-called 'law of the heart' is that it couldn't explain how blood returned from the veins to the right side of the heart.

In the early 1980s physiologists discovered negative intraventricular pressures, i.e., a suction force, in the early diastolic phase indicating that diastole was not a period of passive relaxation but, instead, a period in which blood was actively drawn forward through the veins into the ventricular chamber [3–6]. In order for the ventricle to pump blood through the arteries it first has to contain blood. A handful of studies later found the presence of spiral flow currents in arteries and veins which can only be explained on the basis of a suctional force [7–20].

By the late 1980s numerous studies had established the primacy of diastole in the cardiac cycle and, intriguingly, found that impaired outward movement of the ventricular and arterial walls, known as 'diastolic dysfunction,' was the defining feature of a wide range of chronic conditions: hypertension, diabetes, obesity, depression, cancers autoimmune diseases, as well as organ failure syndromes like chronic kidney disease and chronic heart failure among others [21, 22]. To date scientists have no satisfying explanation for this clustering. The outward motion of the heart and arterial walls, on the other hand, can only be explained on the basis of a mechanical force.

During its cycles of contraction and dilation the heart generates a large magnetic field which is responsible for diastolic expansion. In a series of earlier articles, we describe mechanisms at play in this phenomenon [23–25]. For over a century it has been recognized that the heart and blood contain large iron stores and, while iron's role in various chemical reactions has been exhaustively detailed, there has been little discussion as to whether



iron might play a broader role.

Equally the question arises as to the function served by nerves that course over the surface of the heart. Cardiologists claim these nerves cause the heart to contract but is this correct? As early as the 2nd century AD Roman physician Galen observed in animal experiments that when the heart was cut out and placed in fluid it continued to dilate and contract, what is called cardiac automaticity. By the same token, transplanted hearts continue to function in recipients even though nerve conduction has been interrupted.

What happens during systolic contraction of the ventricle is identical to what happens during the induction of an external magnetic field by electrification of ferrous objects. As the ventricle contracts and iron stores are brought into closer apposition iron nuclei in the heart muscle and blood align and precess synchronously on the basis of field interactions. The flow of electrical currents through nerves saturates the field and induces formation of an external three-dimensional magnetic field within the ventricular chamber causing its expansion. A similar mechanism is at play with magnetic resonance imaging (MRI) in which images are generated by saturation of a magnetic field with radiofrequency pulses.

It cannot be said that recognition of this organized energy field generated by the motions of the heart constitutes a new discovery. Such dynamics were first described by Roman physician Galen around 200 AD and accepted as fact by physicians for over 1500 years until chemically oriented scientists in the 17th and 18th centuries discarded the concept without ever disproving it. Galen's system of humoral medicine was premised on the existence of a blood-borne energy field that gave rise to all bodily functions [26]. Medicine, it seems, has come full circle back to its roots.

While medical scientists acknowledge the centrality of active dilation in cardiovascular dynamics, they advanced the concept of 'endothelial-dependent dilation' to explain such phenomena. They claim that intra-arterial pressure and blood flow induce synthesis and release of the free radical substance nitric oxide which is responsible for vascular dilation. While this may be factually correct it sidesteps the phenomenon of active energy generation, the most significant causal event in the economy of living bodies. To ascribe energy and mechanical forces to molecular causes is like trying to ascribe a thunderstorm to cloud formation: it

is a necessary but not sufficient condition.

COVID-19 Energy Deficiency

One of the most unexpected revelations related to COVID-19 infection has been the degree to which the cardiovascular system is involved in its pathogenesis. Studies indicate that SARS-CoV-2 infects vascular endothelial cells early in the course of the illness leading to inflammation, i.e., endothelitis, which, in advanced cases, extends diffusely throughout the circulatory system. The inflammatory state, in turn, induces further diastolic dysfunction and impairment of energy generation. Some researchers have thus questioned whether the cardiovascular system plays the primary role in mediating the COVID-19 syndrome [27-34].

Endothelial cells form the boundary between blood and the vascular wall and orchestrate energy-dependent processes like smooth muscle contraction and elongation, vessel permeability, coagulation and fibrinolysis. Diastolic and endothelial dysfunction is widely believed to not only impair organ perfusion but augment the prothrombotic state resulting in formation of large and small clots throughout arterial and venous channels.

The ubiquitous distribution of the vascular system explains the diverse range of symptoms and functional disturbances from person to person with apparent random involvement of organs like the lungs, heart, kidneys and brain [35-46]. As indicated above, diastolic dysfunction is the common link among such diverse states as old age, obesity, hypertension, diabetes, chronic heart and kidney disease, all of which increase the risk for severe COVID-19 and mortality. This is to say that pre-existing diastolic dysfunction is the leading prognosticator for poor outcomes.

Diffuse endothelial inflammation in large and small vessels points to a more than coincidental relationship between inflammation and impaired energy-generation. Inflammation is an adaptive cellular response to deficient energy flow across the cell membrane. Diminished intracellular energy induces mitochondrial dysfunction with a shift from aerobic to less efficient metabolic pathways resulting in generation of reactive oxygen species, accumulation of acidic by-products and altered voltage potentials across intracellular membranes [47–53].



Reactive oxygen species cause structural damage by inducing denaturation of proteins, i.e., protein misfolding, as well as formation of the stress-related structure known as the NLRP3 inflammasome which initiates the cytokine storm that accompanies inflammation in COVID-19. Numerous studies link the cytokine storm to both COVID-19 severity and higher mortality rates [54-67]. Blood analysis of COVID-19-infected patients has shown increased TNF- α and pro-inflammatory cytokines including IL-1 β , IL-2, IL-6, and IL-10 which amplify already existing endothelial dysfunction. There is not one but two storms, the cytokine storm and a primary, equally impactful reactive oxygen species storm, which inflicts widespread damage upon intracellular proteins [68-76].

For decades clinicians have speculated on a possible relationship between viral infection and subsequent development of autoimmune disease. This association has come to the forefront in the SARS-CoV-2 pandemic with numerous reports of viral-induced effects mimicking various autoimmune syndromes [77-86]. In both cases pathologic events are associated with altered protein dynamics. The common link is seen in phenomena like autoantibodies (autoAbs), NLRP3 inflammasome formation and neutrophil extracellular traps (NETS).

COVID-19 patients have marked increases in autoAb levels compared with non-infected individuals. As with autoimmune disease, an array of autoAbs have been found directed against endogenous cytokines, chemokines, cell surface proteins as well as RNA and DNA. AutoAbs, depending on the type, may be found in 10%-50% of COVID-19 patients. Since the mid-20th century scientists have claimed that autoAbs were a result of spontaneous genetic mutations that gave rise to 'forbidden clones' of autoAb-producing lymphocytes but evidence surfacing during the pandemic challenges this notion. Studies find that autoAb levels track directly with rising levels of antibodies against SARS-CoV-2 and with disease severity suggesting they form spontaneously during the course of the illness. It is more likely that autoAbs result from protein misfolding related to energy deficiency which, in turn, leads to loss of antigenic specificity and crossreactions with native structures [87-97].

The NLRP3 inflammasome is a complex protein aggregate that forms in the cytoplasm secondary to impaired energy generation by mitochondria. In response to oxidative stress the inflammasome releases pro-inflammatory cytokines into the extracellular fluid (ECF) space initiating

the cytokine storm and, as a coup de grâce, activates cell death (apoptosis) pathways. The NLRP3 complex is found in a host of inflammatory states including autoimmune disorders, Alzheimer's disease, diabetes and atherosclerosis [98–105].

In further support of the energy hypothesis, studies indicate that inflammasome formation is directly related to diastolic and endothelial dysfunction [106-110]. Inflammasomes likely represent the energy-depleted state of a normally folded protein, much like the ventricle at the end of systolic contraction. In the case of the inflammasome, however, energy repletion (repolarization) cannot occur and, instead, the proteins undergo spontaneous aggregation with activation of cell death pathways [111].

Neutrophils (PMNs) are phagocytic cells capable of assimilating and digesting both endogenous and foreign materials. Under conditions of energy depletion, i.e., oxidative stress, the digestive capacity of phagocytic cells is impaired and, as a result, denatured biomolecules accumulate both in cells and in the ECF space. NET release typically occurs during PMN cell death. NETs are large web-like structures containing materials like DNA and a variety of proteins that have spilled into the ECF space following cell injury and death. NETs likely represent energy-depleted proteins that undergo spontaneous aggregation. NETs, found in a variety of autoimmune and inflammatory disorders, are abundant in COVID-19 patients [112-129].

Some scientists claim the purpose of NETs is to trap extracellular materials like bacteria and viruses but this doesn't make sense. NETs, especially DNA, are highly proinflammatory and, instead, likely induce conformational change and aggregation of biomolecules in the ECF space. In addition to triggering autoAb formation, NETS serve as scaffolding for thrombus formation and their presence increases the risk for lung injury, multi-organ damage, and mortality in COVID-19 disease. Formation and accumulation of NETs appears to be primarily due to failure of clearance mechanisms by functionally impaired phagocytic cells.

In the early 1950s cell biologist Christian de Duve described an intracellular membrane-bound organelle he called the lysosome. Later, under the electron microscope, he observed delivery of cellular materials into lysosomes and coined the term autophagy, meaning 'self-eating,' to



designate intracellular digestion [130, 131]. In autophagy acid is concentrated in lysosomes and catabolic enzymes activated not unlike that which occurs in the stomach. Now widely recognized as a function critical to cellular homeostasis, autophagy culls aging and damaged cell structures as well as generating energy through auxiliary pathways during periods of nutrient deficiency [132]. Such orchestrated body-wide autophagic activities constitute what we call the internal digestive system.

As concentration of acid within lysosomal membranes is energy-dependent, impaired mitochondrial function inevitably leads to lysosomal dysfunction and diminished breakdown of defunct cellular structures thus leading to accumulation of undigested material inside and outside of cells. While autophagy plays a key role in all cells, it forms the raison d'être for phagocytic cells of the immune system and thus during periods of energy depletion, such as in advanced COVID-19 infection, deterioration of phagocytic functions is common [133-138].

Evidence we have presented thus points to widespread disruptions in energy generation originating in the cardiovascular system as a hallmark of COVID-19 infection which, secondarily, impairs mitochondrial function and intracellular energy production. Intracellular energy deficits, in turn, induce disturbances in protein metabolism and function, namely, impaired conformational change, i.e., depolarization repolarization- of normally folded proteins and/or misfolding in newly-synthesized proteins. A related aspect of energy depletion and inflammation, accumulation of intra- and extracellular deposits due to impaired autophagy, can also be ascribed to similar origins.

Proteomics

The discovery of intravascular amyloid in COVID-19 patients ties into one of the most protracted and circuitous scientific investigations in search of causality, one that stretches from the early years of the 20th century to the present, and still has yet to fully resolve. The Alzheimer's saga vividly capsulizes the shortcomings of the experimental method.

In 1907 Alöis Alzheimer reported the case of a 51-year-old demented woman at a local asylum in Frankfurt, Germany, constituting the first known description of the disease. Symptoms included loss of memory for recent events along with a cluster of verbal and visual impairments.

After the woman died Alzheimer examined her brain microscopically and observed the typical amyloid deposits and neurofibrillary tangles [139]. Thereafter aggregates became the subject of on-again, off-again investigations but, from the onset, scientists were unable to agree on whether they were cause or effect. Much of the 20th century was spent chasing leads that went nowhere. And yet the solution to the dilemma had been articulated in the early decades of the 20th century but roundly ignored.

In 1984 pathologist George Glenner isolated amyloid-β (Aβ) and showed that it was derived from a cell membrane protein later called amyloid precursor protein (APP) [140]. With little additional evidence Glenner seized on the notion that AB was the cause of Alzheimer's disease (AD). In 1991 a mutation in the gene that codes for APP was discovered in individuals with the familial form of AD leading geneticist John Hardy to advance the amyloid hypothesis imputing that AB deposits represent the primary disease pathology [141]. He argued that Aβ fibrils gradually coalesce into larger strands which morph into the characteristic sheet-like plaques. Such plaques were claimed to trigger pathologic sequelae like synaptic dysfunction, neurofibrillary tangles, inflammation and cell death, all of which invariably progressed to dementia [142].

From the beginning the amyloid hypothesis was on thin ice. Not only did the presence of A β plaques not correlate well with cognitive impairment in AD but geneticists were left trying to explain the presence of plaques in the brains of cognitively normal elderly adults who had died from other causes. Equally problematic was how such aggregates produced cellular injury. Originally it was assumed that plaques were toxic but given that they are insoluble and non-reactive the basis for such toxicity was never established [143].

Most cases of AD are not familial but, instead, spontaneous and not associated with mutations involving APP [144-146]. Dozens of genetic markers have been associated with AD (as well as the other neurodegenerative diseases) but almost none involve precursor proteins to the characteristic deposits. Instead, gene analysis points squarely to impaired autophagy and associated protein degradation pathways [147-153]. Geneticists assumed that since genes code for protein synthesis all protein disorders must have a genetic origin. But protein folding and misfolding is a cytoplasmic function dependent on energy availability.



Proteins possess a complex three-dimensional arrangement. Primary structure, which is genetically coded, refers to the sequence of amino acids that forms the backbone of the chain. Secondary structure comprises a regularly repeating pattern such as the α -helix characteristic of DNA or the β -sheet form typical of amyloid. Such secondary attributes are generated by intermolecular forces like hydrogen bonds and dipole-dipole interactions. The tertiary structure represents the overall 3D conformation of the protein which also is stabilized by non-covalent intermolecular forces [154–157].

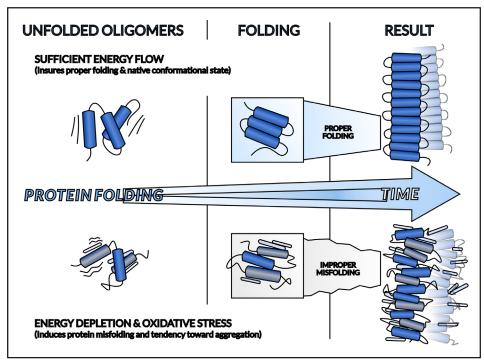
Tertiary structure, known as the native fold, is responsible for the functional properties of a protein. While effecting their functions proteins continually undergo spatial rearrangement, alternating between related conformations as seen, for example, in contraction and dilation of heart muscle. Such changes represent transitions between discrete energy states. In the cardiac cycle, for example, systole represents a period of energy discharge (depolarization) while diastole represents the phase of energy repletion (repolarization).

During or after synthesis most proteins are converted into compactly folded 3D structures many of which are astonishingly complex. Living organisms possess elaborate mechanisms by which to ensure proper folding.

Under certain conditions, as in the congelation of egg white by heat, for example, even normally folded proteins may revert to a misfolded state. By whatever mechanism misfolding occurs, such conformational alterations diminish function. Because of the import of maintaining native conformation, intracellular processes exist to either refold misfolded proteins or to break them into smaller parts for either reassembly or elimination.

The majority of proteins fold in the cytoplasm or endoplasmic reticulum. Both compartments provide not only a proper folding milieu but quality control mechanisms by which to maintain natively folded proteins. A specialized class of macromolecules. i.e., chaperones, discriminate between native and nonnative conformations. If misfolded proteins cannot be properly refolded, they are earmarked for degradation, i.e., intracellular digestion, through a second set of processes known as the ubiquitin-proteasome system. During periods of energy depletion and oxidative stress coordination between various internal processes is impaired and misfolding more likely to occur. Once a critical concentration of misfolded protein is reached, they become prone to the kind of aggregate and inclusion body formation typical of the neurodegenerative disorders and now recognized to play a role in COVID-19 infection [158-162] (Figure 1).

Figure 1. Proper 3D conformation of a protein is dependent on available energy in the cellular milieu. Protein misfolding is more likely to occur during periods of impaired mitochondrial function and oxidative stress.



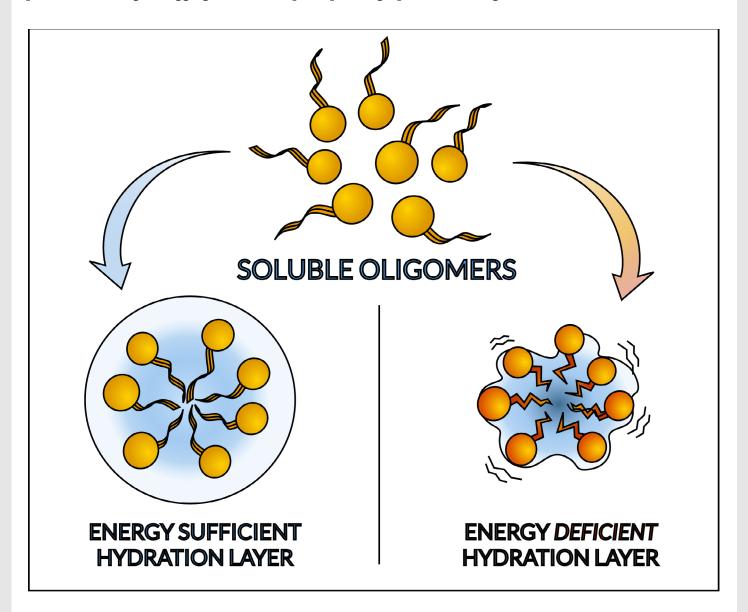


Based on unknown factors amyloid aggregates assume a common secondary structure, the β -sheet pattern, which likely represents a common energetically stable form accessible by a wide range of polypeptides independent of amino acid sequence. This provision is critical since most of the disease related proteins do not share obvious sequence homologies in their native state [163].

Whatever mechanism drives the formation of the β -sheet architecture it is clear that aggregates develop from a series of intermediates beginning with smaller chains,

i.e., oligomers, that gradually self-assemble into larger units. A key feature in the aggregation process is the transition from water-soluble oligomers to insoluble deposits. In the late 1980s pathologists described a peripheral halo surrounding A β aggregates [164]. More recently such halos have been shown to contain soluble A β oligomers suggesting that aggregation bears resemblance to a crystallization process and is driven primarily by dynamics in the surrounding fluid medium [165] (Figure 2). Such recognition opens the door to an entirely new concept of energy flow in living bodies.

Figure 2. Proper 3D conformation is dependent on the expansionary magnetic component of the energy field which enables component domains and strands to repel one another. Oxidative stress, conversely, with its generation of acid by-products, impairs magnetic field strength and augments the contractionary force. This, in turn, leads to protein misfolding and aggregation of susceptible pathologic proteins and oligomers.





Water Dynamics

As impactful as the revolution in proteomics has been it could not have occurred without an equally profound transformation in the conception of water. The recognition of dozens of anomalous properties of water not explainable on the basis of its chemical properties now challenges conventional notions as to its true nature. Emerging evidence substantiates ancient notions of water as a distinct element. Perhaps the most significant contributions have come from biologist Gerald Pollack whose work, *The Fourth Phase of Water*, has triggered yet another decisive paradigm shift [166].

Most significant among the so-called anomalous behaviors of water is its ability to undergo phase transitions between three distinct states—ice, liquid, and vapor—each reflective of its energy content. In experiments Pollack observed that water often underwent spontaneous reorganization and formed a clear zone of variable thickness along the surface of many objects with which it came into contact. Such behaviors had been described by other scientists as early as 1949 but never investigated further. Based on such properties Pollack called the clear layer 'exclusion zone' (EZ) water based upon its tendency to repel solute particles into the adjacent fluid which, in turn, he called 'bulk water'.

Pollack's team found that the EZ tended to form along hydrophilic surfaces and required a molecular template. This explains the peripheral halo researchers observed surrounding Aβ oligomers. Using microelectrodes Pollack et al found significant differences between EZ and bulk water: the EZ was negatively-charged, more alkaline, dense and viscous; bulk water was positively-charged, acidic, with pH often as low as 1-2 suggesting accumulation of protons. As the two phases of water formed, a charge separation took place. The presence of current flow suggested to Pollack that water functions like a battery to generate and conduct energy.

His team deduced that as the EZ acquired spatial order (and physical force) it pushed solute particles into the adjacent bulk water similar to how glaciers extrude rocks. Studies by physical scientists suggest that EZ water has a quasi-crystalline structure and arranges itself in stacked honeycomb sheets, hence the term 'structured water'. Pollack regards it as a distinct fourth phase beyond the traditional solid, liquid and vapor states. As the battery

metaphor implies, structured water appears to play a central role in energy flow. Evidence suggests the fourth phase of water represents a resonant energy state.

In recent decades there has been an explosion of research in the physical sciences using x-ray scattering, NMR spectroscopy, and x-ray crystallography to study protein structure and folding dynamics all of which substantiate the vital role water plays in proteomics [167–171]. To be biologically active proteins must acquire a so-called hydration shell consisting of multiple layers of water molecules sometimes extending up to 25Å from the protein surface. The hydration shell appears to be instrumental in determining not only 3D protein structure but the folding process itself.

The hydration shell, which surrounds hydrophilic domains of most intracellular proteins and membranes, spontaneously organizes into a complex hexagonal lattice-type arrangement, which researchers have compared to a semi-crystalline state. The protein-water complex spontaneously develops structure and assumes gel-like consistency. Hydration shell water surrounding proteins, like the EZ, has physical properties distinct from that of bulk water in the adjacent fluid spaces. Such changes in water state, both in the intracellular and extracellular fluid spaces, would seem to provide an ideal energy source for the many protein-mediated biological processes.

The conjoined role of proteins and water in the energy economy of the body formed a recurrent motif throughout the 20th century. Swedish physical chemist Svante Arrhenius published the first work on the electrical conductivity of ionic solutions, *Investigations on the Galvanic Conductivity of Electrolytes*, in 1894 [172]. He noted that salts dissolved in water split into electrically positive and negative ions that transmit electrical currents in the fluid medium. Based on its tendency to undergo polarization into two opposing species, we refer to this water-mediated energetic component as the dielectric field [173]. Arrhenius conducted studies showing the biological importance of electrolyte dynamics in the interactions between antigens and antibodies.

Arrhenius' work formed the basis for what became known as 'colloid theory' which, simply stated, asserted that ion-containing intracellular water interacts with cell proteins to produce complex three-dimensional structures which, under specific conditions, assume gel-like consistency. In



that bound water conducts electrical currents, changes in energy flux through cells induce conformational changes in the colloid matrix resulting in 'work,' i.e., functional activities like muscle contraction or glandular secretion.

Viennese physician and immunologist Karl Landsteiner, discoverer of the ABO blood groups, was an early proponent of colloid theory. He argued that colloid dynamics and electrochemical forces mediated antigenantibody interactions. It seems, he wrote, 'that this extraordinary type of reaction plays a particularly large part in living organisms; living substance is mostly made up of colloids' [174]. Landsteiner and co-workers found that charged acidic and basic colloids not only moved in opposite directions in electrolysis experiments but precipitated each other. Interactions were based less on chemical constitution than on physical phenomena like pH, solubility and temperature.

Beginning in the 1950s cell biologist Gilbert Ling emphasized the central role of water and colloid dynamics in all cell functions. The cytoplasm, Ling argued, is an integrated system of proteins, water, ions and molecules like ATP that drive all functional processes in the body. All structural elements are linked together by electromagnetic interactions, what he called 'ferromagnetic cooperativity' [175].

In 1962 Ling advanced the association-induction hypothesis based on the notion that electrical polarizations and depolarizations, i.e., induction, were at play in such conformational dynamics [94]. Closecontact interactions among protein chains link them into an organized nexus with secondary, tertiary and even quaternary structure. All colloids exist in open and closed states and undergo reversible transitions based on energy flux. In the years following introduction of his theory Ling's lab was forced to shut down due to inability to secure research grants from funding agencies like the National Institutes of Health.

A recurrent question concerns the nature of the processes that drive protein folding: do amino acid chains randomly generate secondary and tertiary conformations or are conformations determined by the energy state? In the late 1960s Christian Anfinsen called attention to the intricacies of protein folding, particularly the phenomenon of reversibility, which necessarily involves transitional kinetic states. Folding, he argued, is related more to the disposition of electromagnetic forces than to covalent chemical bonding [176].

Cyrus Levinthal pointed out the sheer improbability of protein chains searching randomly through an infinitely large pool of potential configurations to attain native fold [177]. The extreme rapidity with which such folding occurs, on the order of milliseconds, implicates preordained folding pathways related to specific energy states. In 1995 Bryngelson et al showed that in various protein families, like lysozyme for example, polypeptides fold into identical 3D conformations despite extreme disparities in amino acid sequence [178]. They argued in favor of a dynamic energy landscape in which proteins are 'funneled' into their functional energy states [179–183].

For much of the 20th century biological causation was held to be determined by processes originating at the molecular and cellular levels. These assertions are unfounded. Protein folding and misfolding, now widely recognized to drive all functional processes in the body, and to be a leading cause of disease including COVID-19 infection, demand the presence of an organized energy landscape.

Even mainstream medicine seems to be wavering in its century-long infatuation with molecular reductionism. Concerning the relationship between A β and AD science writer Simon Makin, in an influential 2018 Nature editorial, raises the critical question 'is it time to look beyond amyloid- β as the root cause of the condition'? Neuroscientist Michael Murphy comments, 'the time to cast a wider net is now—we need a bigger base of ideas' [184]. Another dramatic paradigm shift is underway.

The Prion Connection

Beyond an association between SARS-CoV-2 infection and amyloid deposition, studies emerging during the pandemic found striking similarities between patterns of COVID-19 propagation and that of classic neurodegenerative diseases like AD and Parkinson's (PD), now recognized to operate on the basis of prion-like mechanisms. Protein misfolding in the nervous system behaves entirely in line with principles we have just established.

The prion diseases are progressive, transmissible neurodegenerative disorders seen in higher vertebrate species: scrapie in sheep, chronic wasting disease in deer, mad cow disease in cattle, and in humans like kuru and Creutzfeldt–Jacob disease [185–187]. The causal agent is a neuronal protein that undergoes pathological misfolding and propagates along nerve tracts in cell–to–cell fashion [188, 189]. Beyond such pathologic descriptions, however,



researchers have never explained exactly how they induce disease. The term prion, rather than denoting a particular disease entity, designates the means by which protein misfolding disorders propagate throughout the nervous system.

During the pandemic it was observed that patients with neurodegenerative disorders like AD and PD were not only more susceptible to severe COVID-19 infection but more likely to experience worsening of their underlying symptoms. Such trends are especially pronounced in AD with multiple reports describing rapid cognitive decline during COVID-19 infection. Moreover, spontaneous cases of AD and PD as well as Creutzfeld-Jacob disease have been described both after SARS-CoV-19 infection as well as COVID-19 vaccination [190-202]. Such phenomena have corollaries at the molecular level.

Injection of purified A β material into AD-prone mice induces accelerated deposition of A β and intracellular aggregation of tau-protein with neurofibrillary tangle formation in surrounding brain tissue of the mice [203-204]. Based on such behaviors it is suggested that A β possesses prion-like behaviors. By the same token, the presence of the SARS-CoV-2 protein in the serum has been found to enhance amyloid formation [205-206]. In laboratory studies the SARS-CoV-2 protein interacts with α -synuclein to induce Lewy body formation in PD-prone cell lines [207]. Other studies find that interactions between the SARS-CoV-2 protein and α -synuclein also accelerate amyloid formation [208]. Such intertwined phenomena constitute what researchers refer to as prion-like transmission.

To explain such behaviors on a cellular and molecular basis scientists describe four attributes of prion-like propagation: intracellular conversion of a natively-folded protein into a misfolded form; extrusion of misfolded protein into the ECF space with uptake by neighboring cells; the ability of misfolded species to induce misfolding in other similar proteins; and the tendency of misfolded proteins to induce misfolding at distant sites [209, 210]. Based on such spurious criteria, a misfolded protein must somehow exit one cell, travel through the ECF space, enter another cell and, by some vague 'template' mechanism, induce protein misfolding in other cells. This is pure science fiction. A far simpler explanation is that introduction of pathogenic protein material into the system places additional stress on degradation pathways with subsequent accumulation of aggregates.

From an energetic standpoint such physical mechanisms aren't necessary. Only three phenomenamust be accounted for in all the related disorders: Why do proteins misfold? Why do they accumulate in cells or the ECF space? And, importantly, what is the basis for this tendency toward aggregation? The aggregation question will be addressed in the final section.

We have already established that deficient energy flow into cells and impaired mitochondrial function are responsible for all of the disturbances. The fact that proteins in nearby cells undergo subsequent misfolding and aggregation can only indicate an expanding energy debt. In that mitochondrial dysfunction and oxidative stress impair lysosomal function, the accumulation of A β oligomers, Lewy bodies, neurofibrillary tangles and other misfolded proteins can only be ascribed to deficient autophagy. Not only is direct contact between proteins not necessary to induce misfolding, there is no plausible mechanism by which this should even occur. Protein conformation is purely related to energy status. End of story. It is surprising that cell biologists have failed to reach consensus on such a critical issue.

It goes without saying that the brain and nervous system do not represent the primary route of infection for SARS-CoV-2. While CNS manifestations such as stroke, hemorrhage, or inflammation can be attributed to the vascular system, other manifestations take origin by a distinctly different route. Of great relevance to the spread of so-called prion disease in the nervous system is recognition of early functional disturbances which point with inerrant accuracy to where the pathogenic action unfolds.

Given the well-established oral route for propagation of various prion diseases, it is not surprising that in recent years both AD and PD have been linked to this mechanism. Early in the course of PD misfolded proteins accumulate in gut lymphoid tissue and spread to organs like the spleen, tonsils, appendix and the enteric nervous system [211-219]. A peripheral origin for PD is supported by the early appearance of autonomic dysfunction [220-224]. Subjects often develop symptoms like decreased saliva production, dysphagia, impaired gastric emptying or constipation. Of particular significance is the presence of disorders of smell in up to 90% of early-stage PD and 85% of early-stage AD patients [225-229]. Such symptoms usually precede the onset of motor disturbances, cognitive decline and dementia.



Based on such evidence it has been suggested that misfolded proteins, acting in prion-like fashion, gain access to the CNS via two routes: one through the nose into the olfactory nerve, and the second through the intestinal mucosa and into the vagus nerve. The nasal route would account for the high percentage of PD subjects with olfactory symptoms and, in more advanced cases, pathology in the forebrain. Retrograde propagation of α -synuclein misfolding along the vagus nerve not only explains various autonomic dysfunctions but involvement of the dorsal motor nucleus in the medulla [230–232].

Surprisingly few reports have commented on the well-documented association between neurodegenerative disorders and autonomic dysfunction [233–236]. Autonomic dysfunction is a frequent accompaniment of all the dementias and includes postural hypotension, dizziness, gastrointestinal disturbances and urinary incontinence. Autonomic symptoms are associated with poorer disease outcomes. Such imbalances indicate altered nerve traffic patterns and trace directly to decreased energy flow in the ECF space which itself is secondary to diminished energy generation by the cardiovascular system.

The same early symptom patterns are observed in subjects with SARS-CoV-2 infection [237-244]. In various studies disorders of smell and taste range from 10% to as high as 85% with a mean in the 60% range. Such chemosensory dysfunction has also been reported following COVID-19 vaccination. Deficits may occur before or coincident with onset of other manifestations. Symptoms are generally self-limited and range from 3-4 days up to several months.

Autonomic dysfunction as detected by heart rate variability testing is a frequent accompaniment of COVID-19 infection. Of special interest is the recognition of widespread autonomic dysfunction in subjects with the long-COVID syndrome. Symptoms may include fatigue, palpitations, tachycardia, chest pain, shortness of breath, orthostatic hypotension, loss of smell and/or taste, exercise intolerance, headaches, 'brain fog' and difficulty concentrating, sleep disturbances as well as depression and/or anxiety [245-255]. The syndrome, believed to affect 10-20% of infected individuals, generally persists for weeks to months. How to reconcile the striking similarities between COVID-19 and early forms of the neurogenerative disorders?

To explain energy flow through nerves, early 20th century scientists posited that neurons functioned like tiny batteries and generated their own electrical currents. But this explanation doesn't fly in the face of dramatic events like sudden cardiac arrest in which there is immediate cessation of all neuronal functions and loss of consciousness the moment cardiac activity ceases. How to account for such a tight functional linkage between the two compartments?

Given the directionality of flow currents in nerves the question arises as to where peripheral sensory nerves, which course toward the brain and spinal axis, derive their currents. The unavoidable conclusion is that they originate in the ECF space just as Pollack's structured water concept would suggest. By the same token, given that the surface of the cerebral hemispheres is lined by dendrites, which convey currents directionally into the deeper brain structures, and which are in direct contact with cerebrospinal fluid, one must draw similar conclusions as to the origin of electrical currents in the hemispheres.

The flow of currents in the nervous system, necessarily, is driven by cardiac dynamics. Each systolic contraction of the heart creates a suctional force in nerves which draws currents forward into both peripheral sensory nerves and dendrites over the outer surface of the brain. When the heart dilates it moves fluids; when its contracts it moves energy currents. This substantiates the claim of Swedish radiologist Björn Nordenström in his pioneering (but overlooked) work Biologically Closed Electric Circuits (1983) that there is not one but two overlapping circulations, one consisting of the flow of fluids and the other of energy currents [256].

Such a model goes a long way in explaining early functional deficits such as loss of smell and taste as well as the plethora of autonomic imbalances that define both acute and long-haul COVID-19 syndromes as well as the early neurodegenerative disorders. It is axiomatic that therapeutic attempts must be aimed at restoration of energy deficits and correction of imbalances.

Aggregation Dynamics

So, we return to the point at which our deliberations began: deposition of amyloid aggregates in the vascular compartment. It is established that accumulation of amyloid fibrils is the end result of mitochondrial



dysfunction and impaired intracellular digestion of misfolded proteins. But another question looms large: why are pathologic species prone to aggregation? What dynamics are in play that would precipitate such events?

Recently, scientists discovered that fibrinogen, precursor to the clot-forming protein fibrin, can, under certain circumstances, transform into a misfolded form of fibrin not unlike the β -sheet rich amyloids and prions [257-259]. This pathologic isoform propagates intravascularly forming micro-clots which, in turn, entrap other proteins and propagate to form larger clots. Such thrombi persist in the vascular system indefinitely and, eventually, may occlude small and large arteries and veins with predictable consequences. It was further observed that such atypical clots are highly resistant to standard anticoagulant therapy and may require prolonged treatment with multiple agents if they first don't cause irreversible organ injury or death.

By the same token, in lab experiments when the SARS-CoV-2 protein was added to the blood it resulted in structural changes not only to fibrinogen but other blood proteins like prothrombin and complement [260-264]. The spike protein binds to a host of aggregation-prone proteins like heparin, the heparin binding proteins, A β , α -synuclein, tau and others to accelerate aggregation of pathological amyloid proteins. Reports describe multiple amyloidogenic and prion-like domains in the spike protein. How can such phenomena be explained? Examination of the prion protein provides further insight.

The prion protein exists in two stable energy-dependent isoforms, i.e., natively folded and misfolded states. The functional domain of the protein consists of three α -helix and two β -sheet regions. In its natively-folded conformation α -helix-rich regions predominate while the pathogenic misfolded state is characterized by β -sheet dominance. This pathologically folded protein, considered 'toxic' by researchers, gives rise to the observed pathology. Breakdown of the misfolded prion

protein generates A β oligomers. Like other misfolded proteins the prion protein tends to aggregate not only with itself but also other proteins [265–273].

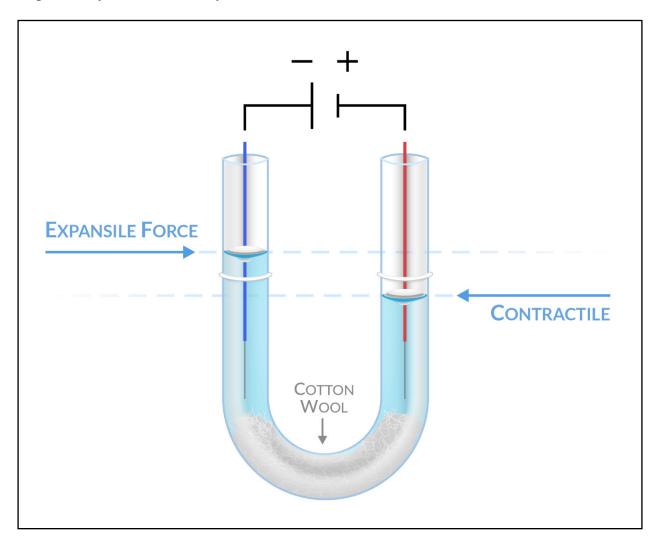
It seems likely that when aggregation prone proteins undergo misfolding they revert to stable conformations like the β-sheet-dominant form which cannot expand fully into their 3D conformations. Such misfolding induces compaction of constituent chains and inability to repel nearby proteins. This mechanism has received increased attention in recent years [274-278]. Crowded physiological environments, both intra- and extracellular, favor protein aggregation by promoting conformational instability, inhibiting breakdown of accelerating seeding of amyloids, and promoting prionlike replication. Such events, in turn, translate into pathological changes in the blood.

During the pandemic studies found that SARS-CoV-2 infected individuals had higher blood viscosity than non-infected persons and, among the infected, higher viscosity measurements translated into higher mortality [279-281]. Higher blood viscosity is associated with diastolic dysfunction, blood flow disturbances, increased shear stress, increased tendency for platelet and red blood cell aggregation, and, finally, increased blood coagulation. Energy deficiency, protein misfolding and water dynamics come full circle in the blood and must be explained on an entirely different basis than simply molecular factors.

In an experiment intended to demonstrate the effects of electric fields in water Nordenström packed a U-shaped glass tube with cotton wool in its lower curved portion to simulate capillary resistance [282] (Figure 3). Both limbs of the tube were filled with water and metallic electrodes connected to a DC power source were placed on each side. After a variable period of time, depending on the size of the tube and strength of the battery, one observes differential water levels in the two limbs with the left (cathodic) side higher than the right (anodic) pole. What is happening here?



Figure 3. A U-shaped glass tube is packed with cotton wool in its lower curved portion and filled with water. Metallic electrodes connected to a DC power source are introduced on each side. After a variable period of time one observes differential water levels in the two limbs with the cathodic side higher than the anodic pole, illustrating the conjoined expansionary and contractionary forces within the dielectric field of the water medium.



Scientists would ascribe such results to electrically-induced migration of charged species, including water, with differential accumulation at the two poles. Such differential levels are no more explainable on the basis of water and ion movement than are the oceanic tides. The spatial reorganization of water is purely a field-mediated effect. Real forces produce differential fluid levels and the migration of ion species.

Electrical currents streaming through the electrode at the cathode generate a magnetic field around the wire which causes expansion of water. By the same token, currents drawn out by the anode induce contraction of the surrounding water which causes its level in the tube to drop. The dielectric field is composed of two opposing forces, the expansionary magnetic and a contractionary or

counter spatial force. When we say water is polarizable, we mean that electrical currents cause the two components of the dielectric field to separate and assert their opposing effects. And as with an automotive battery aggregates tend to precipitate out at the anodic pole where the contractionary force is strongest.

We can thus say with certainty that protein misfolding and aggregation reflect the energy content of the milieu in which they reside. In the cardiovascular system, as we have shown, the outward movement of the ventricular and arterial walls is related to generation of a magnetic force. Magnetism is the only energy form possessing 3D spatiality. As the field weakens, as in diastolic dysfunction, the contractionary force gains strength thereby altering fluid dynamics within both in the



intravascular compartment and ECF giving rise to downstream pathologic carnage. As Galen's functionally based humoral system of medicine asserts, *quae incipit in sanguine:* everything begins in the blood.

Without a doubt such dynamics are in play during COVID-19 infections and account not only for protein misfolding and aggregation in the blood, resulting in atypical thrombi resistant to anticoagulant therapy, but in the peripheral nervous system giving rise to symptoms associated with autonomic dysfunction. Increasing evidence links such phenomena to the rising tide of long-COVID-19 cases [283-285]. Misfolding dynamics also explain *in toto* the proliferation of autoantibodies associated with primary SARS-CoV-2 infection as well as the COVID-19 vaccines, as seen in the well-described vaccine induced thrombotic thrombocytopenia (VITT)

syndrome. None of the therapies currently employed by medical science effectively addresses a single one of these energy-related disturbances. As we pointed out in a previous piece, 80-90% of the COVID-19 pandemic-related deaths were preventable had appropriate measures been implemented.

We spoke with an embalmer, one who preserves bodies for ceremonial burial practices, who claims that since introduction of the vaccines he has encountered mortuarial phenomena he had never seen before in his career. After the COVID-19 pandemic and introduction of mRNA vaccines he began to encounter many cases with extensive arterial and venous clot formation. Other embalmers in the US and Europe describe similar phenomena. He sent pictures of clots he extracted from decedents (Figure 4).

Figure 4. Variegated post-mortem clots typical of those embalmers have encountered with increasing frequency in the COVID-19 era. (Courtesy of Richard Hirschman)

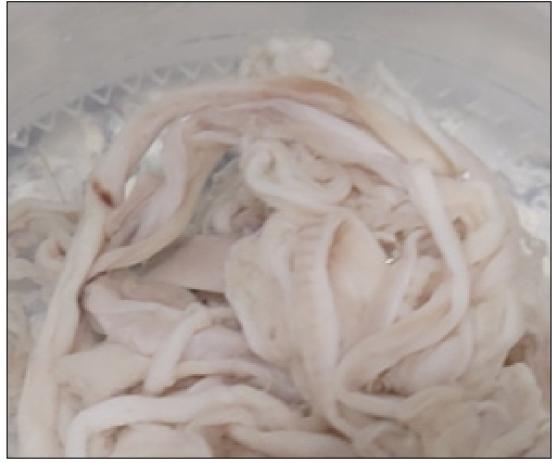




Assembly by Steve Barshov











A colleague of ours who is in robust health received two doses of the Pfizer vaccine in April 2021. Five months later, for no apparent reason, he developed extensive lower extremity deep vein thrombosis. After months of anticoagulant treatment, he continued to experience intermittent swelling and pain. How many latent arterial and venous thromboses, autoimmune phenomena, and chronic conditions like AD and PD are insidiously propagating in post-COVID-19-infected subjects or in those who received the vaccine? This looming catastrophe is all the more disturbing given that medical scientists mishandled the pandemic from the onset and have only magnified its detrimental impact.

References

- 1. The Structure of Scientific Revolutions. Thomas. Kuhn University of Chicago Press, 1962
- 2. Anatomical Studies on the Motion of the Heart and Blood (1628). William Harvey. Leaketranslation; Charles Thomas Publisher, Springfield Illinois, 1958
- 3. Negative intraventricular diastolic pressure in patients with mitral stenosis: evidence of left ventricular diastolic suction. Sabbah HN, Anbe DT, Stein PD. *Am J Cardiol* 1980; 45(3): pp 562-66.
- 4. Left ventricular diastolic suction as a mechanism

- of ventricular filling. Hori M, Yellin EL, Sonnenblick EH. *Jpn Circ J* 46(1): pp 124-129; 1982
- 5. The heart as a suction pump. Robinson TF, Factor SM, Sonnenblick EH. *Scientific American* 254(6): pp 84-91;1986
- 6. The heart is not a pump: A refutation of the pressure propulsion premise of heart function. Marinelli R, Fürst B, van der Zee H, McGinn H, Marinelli W. *Frontier Perspectives* 5(1): pp 15-24;Fall-Winter 1995
- 7. Spiral laminar flow in arteries? Stonebridge PA, Brophy CM. *Lancet* 338(8779):pp 1360-61;1991
- 8. Spiral laminar flow in vivo. Stonebridge PA, Hoskins PR, Allan PL, Belck JF. *Clin Sci (Lond)*91(1): pp 17-21;1996
- 9. Helical and retrograde secondary flow patterns in the aortic arch studied by three-directional magnetic resonance velocity mapping.Kilner PJ, Yang GZ, Mohiaddin RH, Firmin DN, Longmore DB. *Circulation*88(5):pp 2235-47;1993
- 10. Physiological significance of helical flow in the arterial system and its potential clinical applications.Liu X, Sun A, Fan Y, Deng X. *Ann Biomed Engineer*43(1): pp 3-15; 2015



- 11. Three-dimensional blood flow dynamics: spiral/helical laminar flow. Stonebridge PA. *Methodist Debakey Cardiovasc J.* 2011; 7(1): 21-26
- 12. Spiral laminar flow: A survey of a three-dimensional arterial flow pattern in a group of volunteers. Stonebridge PA, Suttie SA, Ross R, Dick J. *Eur J VascEndovasc Surg.* 2016; 52(5): 674-80
- 13. Patterns of flow in the left coronary artery. Sabbah HN, Walburn FJ, Stein PD. *J BiomechEngin*1984;106(3): 272-79
- 14. Flow visualization study of spiral flow in the aorta-renal bifurcation. Fulker D, Javadzadegan A, Li Z, Barber T. *Comput Meth Biomech Biomed Engin*2017; 20(13): 1438-41
- 15. The mechanics of spiral flow: Enhanced washout and transport. Huang Zhang P, Tkatch C, Newman R, Grimme W, et al. *Artif Organs*2019; 43(12): 1144-53
- 16. Parallel and spiral flow patterns of vertebral artery contributions to the basilar artery. Smith AS, Belton JR. *Am J Neuroradiol*1995; 16(8): 272-79
- 17. Flow patterns in the human carotid artery bifurcation. Motomiya M, Karino T. *Stroke*1984;15(1): 50-56
- 18. Spiral systolic blood flow in the ascending aorta and aortic arch analyzed byechodynamography. Tanaka M, Sakamoto T, Sugawara S, Nakajima H, et al. *J Cardiol*2010; 56(1): 97-110
- 19. Flow patterns in dog aortic arch under a steady flow condition simulating mid-systole. EndoS, Sohara Y, Karino T. *Heart Vessels* 1996; 11(4): 180-91
- 20. Blood flow analysis of the aortic arch using computational fluid dynamics. Numata S, Itatani K, Kanda K, Doi K, et al. *Eur J Cardiothorac Surg*2016; 49(6): 1578-85
- 21. Diastolic Dysfunction. Little WC, Cheng CP. *Cardiol Rev*6(4): pp 231-239;1988
- 22. State of the art: 'diastology' research 1998.Oki T. *J Med Invest* 45(1-4): pp 9-25;1998
- 23. Aether, fields & energy dynamics in living bodies Part I.Thorp KE, Thorp JA, Walker PR. *G Med Sci*2021; 2(5): 014-025.
- 24. Aether, fields & energy dynamics in living bod-

- ies Part II. Thorp KE, Thorp JA, Walker PR. *G Med Sci*2021; 2(6): 001-020.
- 25. Aether, fields & energy dynamics in living bodies Part III. Thorp KE, Thorp JA, Walker PR. *G Med Sci*2021; 2(6): 021-047.
- 26. *Galen's System of Physiology and Medicine*. Rudolph E. Siegel publ.S. Karger, Basel, Switzerland 1968; pp. 30-47
- 27. The vascular endothelium: the cornerstone of organ dysfunction in severe SARS-CoV-2 infection.Pons S, Fodil S, Azoulay E, et al. *Crit Care*2020;24(1):353
- 28. Understanding COVID-19: in the end is it endothelium—what else? Lüscher TF. *Eur Heart J* 2020;41(32):3023-27
- 29. COVID-19 is, in the end, an endothelial disease. Libby P, Lüscher T. *Eur Heart J* 2020;41(32):3038-44
- 30. Endothelial dysfunction in COVID-19: lessons learned from coronaviruses. Gavriilaki E,Anyfanti P, Gavriilaki M, et al. *CurrHypertens Rep*2020; 22(9):63
- 31. Endothelial dysfunction in COVID-19: A position paper of the ESC Working Group forAtherosclerosis and the ESC Council of Basic Cardiovascular Science. Evans PC, Rainger GE, Mason JC, et al. *Cardiovasc Res*116(14):2177-84
- 32. Endothelial cells and SARs-CoV-2: An intimate relationship.Barbosa LC, Gonçalves TL, deAraujo LP, et al. *VascPharmacol*2021; 137:106829
- 33. Endothelial dysfunction in COVID-19: Current findings and therapeutic implications. NägeleMP, Haubner B, Tanner FC, et al. *Atherosclerosis*2020;314:58-62
- 34. COVID-19 and cardiovascular consequences: Is the endothelial dysfunction the hardestchallenge?Del Turco S, Vianello A, Ragusa R, et al. *Thromb Res*2020;196:143-51
- 35. COVID-19, the pandemic of the century and its impact on cardiovascular diseases. Zhang Y, Wang M, Zhang X, et al. *CardiolDiscov*2021; 1(4):233-58
- 36. Cardiovascular manifestations of COVID-19 infection.Magadum A, Kishore R. *Cells*2020;9(11):2508



- 37. COVID-19 and the cardiovascular: A comprehensive review. Azevedo RB, Botelho BG,Hollanda JVG, et al. *J Hum Hypertens*2021; 35(1):4-11
- 38. Low brachial flow-mediated dilation predicts worse prognosis in hospitalized patients with CO-VID-19. Bianconi V, Mannarino MR, Figorilli F, et al. *J Clin Med*2021; 10(22):5456
- 39. Immunity, endothelial injury and complement-induced coagulopathy in COVID-19.Perico L,Benigni A, Casiraghi F, et al. *Nat Rev Nephrol*2021; 17(1):46-64
- 40. Endothelial dysfnction and thrombosis in patients with COVID-19: Brief report.NagashimaS, Mendes MC, Camargo Martins AP, et al. *AtherosclerThromb-Vasc Biol* 2020; 40(10):2404-07
- 41. Endothelial activation and dysfunction in COV-ID-19: From basic mechanisms to potentialtherapeutic approaches. Jin Y, Ji W, Yang H, et al. *Signal Transduct Target Ther*2020;5(1):293
- 42. Inflammation resolution: A dual-pronged approach to averting cytokine storms in COVID-19? Panigrahy D, Gilligan MM, Huang S, et al. *Cancer Metastasis Rev*2020;39(2):337-40
- 43. COVID-19 may predispose to thrombosis by affecting both vascular endothelium andplatelets. Cure E, Cure MC. *Clin Appl Thromb Hemost*2020; 26:1076029620933945
- 44. Endothelial dysfunction contributes to COVID-19-associated vascular inflammation and coagulopathy. Zhang J, Tecson KM, McCullough PA. *Rev Cardiovasc Med*2020;21(3):315-19
- 45. The coagulopathy, endotheliopathy, and vasculitis of COVID-19. Iba T, Connors JM, Levy JH. *Inflamm Res* 2020; 69(12):1181-89
- 46. COVID-19-associated coagulopathy and disseminated intravascular coagulation. Asakura H, Ogawa H. *Int J Hematol* 2021;113(1):45-57
- 47. COVID-19 and ROS storm: What is the forecast for hypertension? de Oliveira AA, Priviero F,Lima VV, et al. *Am J Hypertens*2021; 34(8):779-82
- 48. Elucidating of oxidative distress in COVID-19 and methods of its prevention. Barciszewska AM. *Chem Biol Interact* 2021; 344:109501

- 49. What can cellular redox, iron, and reactive oxygen species suggest about the mechanisms and potential therapy of COVID-19. Muhoberac BB. *FrontCell Infect Microbiol* 2020; 10:569709
- 50. Oxidative stress as key player in Severe Acute Respiratory Coronavirus (SARS-CoV) infection.Delgado-Roche L, Mesta F. *Arch Med Res*2020;51(5):384-87
- 51. Neurological implications of COVID-19: role of redox imbalance and mitochondrialdysfunction. Kaundal RK, Kalvala AK, Kumar A. *Molecular Neurobiol*2021; 58(9):4575-87
- 52. Reactive oxygen species, proinflammatory and immunosppressive mediators induced in COVID-19: overlapping biology with cancer. Kalyanaraman B. *RSC Chem Biol*2021;2(5):1402-14
- 53. Tissue damage damage from neutrophil-induced oxidative stress in COVID-19. Schönrich G, Raftery MJ, Samstag Y, et al. *Nat Rev Immunol*2020; 20(9):515-16
- 54. The longitudinal immune response to coronavirus disease 2019: Chasing the cytokine storm. Chau AS, Weber AG, Maria NI, et al. *Arthritis Rheumatol*2021; 73(1):23-35
- 55. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. Huang C, Wang Y, Li X. *Lancet*2020 Feb 15;395(10223):497-506.
- 56. Presenting Characteristics, Comorbidities, and Outcomes Among 5700 Patients Hospitalized with COVID-19 in the New York City Area. Richardson S, Hirsch JS, Narasimham M. *JAMA*2020;323(2):2052-59
- 57. Clinical course and risk factors for mortality of adult inpatients with COVID-19 in Wuhan, China: A retrospective cohort study. Zhou F, Yu T, Du R, et al. *Lancet* 2020; 395(10229):1054-62
- 58. Clinical characteristics and survival analysis in critical and non-critical patients with COVID-19 in Wuhan, China: A single-center retrospective case control study. Tian R, Wu W, Wang C, et al. *Sci Rep*2020; 10(1):17524
- 59. Haematological characteristics and survival analysis in the classification and prognosisevaluation of CO-VID-19: A retrospective cohort study. Liao D, Zhou F, Luo L, et al. 2020;7(9):e671-78



- 60. Factors associated with death outcome in patients with severe coronavirus disease-19(COVID-19): A case-control study. Pan F, Yang L, Li Y. *J Med Sci*2020; 17(9):1281-92
- 61. Clinical characteristics of patients with severe pneumonia caused by SARS-CoV-2 in Wuhan, China. Wang Y, Zhou Y, Yang Z, et al. *Respiration*2020; 99(8):649-57
- 62. COVID-19: consider cytokine storm syndromes and immunosuppression.Mehta P, McAuleyDF, Brown M, et al. *Lancet* 2020;395(10229):1033-34
- 63. Clinical predictors of mortality due to COVID-19 based on an analysis of data of 150 patients from Wuhan, China.Ruan Q, Yang K, Wang W, et al. *J Intensive Care Med* 2020; 46(5):846-48
- 64. Predictors of COVID-19 severity: A literature review.Gallo Marin B, Aghagoli G, Lavine K, etal. *Rev Med Virol* 2021; 31(1):1-10
- 65. Clinical characteristics and day-90 outcomes of 4244 critically ill adults with COVID-19: A prospective cohort study. COVID-ICU Group on behalf of the REVA Network and the COVID-ICU. *Intensive Care Med* 2021;47(1):60-73
- 66. Cytokine storm in COVID-19: Pathogenesis and overview of anti-inflammatory agents used in treatment. Soy M, Keser G, Atagündüz P, et al. *Clin Rheumatol* 2020; 39(7):2085-94
- 67. Controlling the cytokine storm is vital in CO-VID-19. Tang L, Yin Z, Hu Y, et al. *Front Immu-nol*2020;11:570993
- 68. Inflammatory response in COVID-19 patients resulting from the interaction of theinflammasome and SARS-CoV-2. Cheon SY, Koo BN. *Int J Mol Sci* 2021;22(15):7914
- 69. Targeting the NLRP3 inflammasome in severe COVID-19. Freeman TL, Swartz TH. *FrontImmunol* 2020;11:1518
- 70. Inflammation and pyroptosis as therapeutic targets for COVID-19. Yap JKY, Moriyama M,Iwasaki A. *J Immunol* 2020; 205(2):307-12
- 71. Controlling the cytokine storm is vital in CO-VID-19.Tang L, Yin Z, Hu Y, et al. *Front Immu*-

- nol2020;11:570993
- 72. The cytokine storm and COVID-19.Hu B, Huang S, Yin L. *J Med Virol* 2021; 93(1):250-56
- 73. Cytokine storm in COVID-19: pathogenesis and overview of anti-inflammatory agents used in treatment.Soy M, Keser G, Atagündüz P, et al. *Clin Rheumatol*2020; 39(7):2085-94
- 74. COVID-19: Consider cytokine storm syndromes and immunosuppression. Mehta P, McAuleyDF, Brown M, et al. *Lancet* 2020; 395(10229):1033-34
- 75. Immunopathology of Hyperinflammation in COVID-19. Gustine JN,Jones D. *Am J Pathol*2021; 191(1):4-17
- 76. Hyperinflammation and immune response generation in COVID-19. Mishra KP, Singh AK,Singh SB. *Neuroimmunomodulation*2020; 27(2):80-86
- 77. Autoantibodies related to systemic autoimmune rheumatic diseases in severely ill patients with COV-ID-19. Vlachoyiannopoulos PG, Magira E, Alexopoulos H, et al. *Ann Rheum Dis*2020; 79: 1661–1663.
- 78. COVID-19 and autoimmunity. Ehrenfeld M, Tincani A, Andreoli L, et al. *Autoimmune Rev*2020; 19(8): 102597
- 79. Seronegative erosive osteoarthritis following SARS-CoV-2 infection. Drosos AA, Pelechas E,Voulgari PV. *RheumatolTher*2021; 16:1-7
- 80. Autoimmune and rheumatic musculoskeletal diseases as a consequence of SARS-CoV-2infection and its treatment. Shah S, Danda D, Kavadichanda C, et al. *Rheumatol Int*2020;40(10): 1539-54
- 81. Concomitant new diagnosis of systemic lupus erythematosus and COVID-19 with possibleantiphospholipid syndrome. Just a coincidence? A case report and review of intertwining pathophysiology. Mantovani Cardoso E, Hundal J, et al. *Clin Rheumatol*2020; 39(9): 2811-2815
- 82. The SARS-CoV-2 as an instrumental trigger of autoimmunity. Dotan A, Muller S, Kanduc D, etal. *Autoimmun Rev* 2021 Apr; 20(4):102792
- 83. SARS-CoV-2 infection as a trigger of autoimmune response.Sacchi MC, Tomiazzo S,Stobbione P, et al. *Clin Transl Sci*2021 May;14(3):898-907



- 84. Autoimmune complications of COVID-19.Yaz-danpanah N, Rezaei N. *J Med Virol*2022 Jan;94(1):54-62
- 85. Autoimmunity is a hallmark of post-COVID syndrome. Rojas M, Rodríguez Y, Acosta-Ampudia Y, et al. *J Transl Med*2022 Mar; 20(1):129
- 86. The intersection of COVID-19 and autoimmunity. Knight JS, Caricchio R, Casanova JL, et al. *JClin Invest*2021 Dec;131(24):e154886
- 87. Diverse functional autoantibodies in patients with COVID-19. Wang EY, Mao T, Klein J, et al. *medRx-iv*2021 Feb:2020.12.10.20247205
- 88. New-onset IgG autoantibodies in hospitalized patients with COVID-19. Chang SE, Feng A,Meng W, et al. *Nat Commun*2021 Sep;12(1):5417
- 89. Autoantibodies neutralizing type I IFNs are present in \sim 4% of uninfected individuals over 70years old and account for \sim 20% of COVID-19 deaths. Bastard P, Gervais A, Le Voyer T, et al. *SciImmunol* 2021;6(62):eabl4340
- 90. Pre-existing autoantibodies neutralizing high concentrations of type I Interferons in almost10% of COVID-19 patients admitted to intensive care in Barcelona. Solanich X, Rigo-Bonnin R, Gumucio VD, et al. *J Clin Immunol*2021 Nov;41(8):1733-44
- 91. Paradoxical sex-specific patterns of autoantibody response to SARS-CoV-2 infection.Liu Y,Ebinger JE, Mostafa R, et al. *J Transl Med*2021 Dec; 19(1):524
- 92. Positive anti-SSA/Ro antibody in a woman with SARS-CoV-2 infection using immunophenotyping: A case report. Huang PI, Lin TC, Liu FC, et al. *Medicina* (*Kaunas*)2020;56(10): 521
- 93. High levels of anti-SSA/Ro antibodies in COV-ID-19 patients with severe respiratory failure: a case-based review: High levels of anti-SSA/Ro antibodies in COVID-19. Fujii H, Tsuji T, Yuba T, Tanaka S, et al. *Clin Rheumatol*2020; 39(11): 3171-75
- 94. Diverse functional autoantibodies in patients with COVID-19. Wang EY, Mao T, Klein J, et al. *medRx-iv*2021; 2020.12.10.20247205
- 95. Prothrombotic autoantibodies in serum from patients hospitalized with COVID-19.Zuo Y,Estes SK, Ali RA, et al. *Sci Transl Med*2020;12(570): eabd3876

- 96. Autoantibodies against type I IFNs in patients with life-threatening COVID-19.Bastard P,Rosen LB, Zhang Q, et al. *Science* 2020; 370(6515):eabd4585
- 97. Antinuclear antibodies (ANAs) detected by indirect immunofluorescence (IIF) method inacute COVID-19 infection: Future roadmap for laboratory diagnosis.Peker BO, Şener AG,KaptanAydoğmuş F. *J Immunol Methods* 2021; 499:113174
- 98. The NLRP3 Inflammasome: An Overview of Mechanisms of Activation and Regulation.KelleyN, Jeltema D, Duan Y, He Y. *Int J Mol Sci* 2019 Jul; 20(13):3328
- 99. Inflammasome activation and assembly at a glance.Malik A, Kanneganti TD. *J Cell Sci*2017 Dec;130(23):3955-63
- 100. Stressing out the mitochondria: mechanistic insights into inflammasome activation. Yabal M, Calleja DJ, Simpson DS, et al. *J Leukocyte Biol* 2019 Feb; 105(2):377-99
- 101. NLRP3 inflammasome in endothelial dysfunction.Bai B, Yang Y, Wang Q, et al. *Cell Death Dis* 2020 Sep; 11(9):776
- 102. NLRP3 inflammasome at the interface of inflammation, endothelial dysfunction, and type IIdiabetes. Gora IM, Ciechanowska A, Ladyzynski P. *Cells* 2021 Feb;10(2):314
- 103. Contribution of redox-dependent activation of endothelial NLRP3 inflammasomes tohyperglycemia-induced endothelial dysfunction. Chen Y, Wang L, Pitzer AL, et al. *J Mol Med(Berlin)* 2016 Dec: 94(12):1335-47
- 104. Structure of the NLRP3 decamer bound to the cytokine release inhibitor CRID3.HochheiserIV, Pilsl M, Hagelueken G, et al. *Nature* 2022 Apr;604(7904):184-89
- 105. The NLRP3 inflammasome: molecular activation and regulation to therapeutics. SwansonKV, Deng M, Ting JP-Y. *Nature Rev Immunol* 2019 Apr; 19:477-89
- 106. Suppression of NLRP3 inflammasome activation ameliorates chronic kidney disease-induced cardiac fibrosis and diastolic dysfunction. Bugyei-Twum A, Abadeh A, Thai K, et al. *Sci Rep* 2016 Dec;6:39551
- 107. Preservation of contractile reserve and diastolic



- function by inhibiting the NLRP3inflammasome with OLT1177 (Dapansutrile) in a mouse model of severe ischemiccardiomyopathy due to non-reperfused anterior wall myocardial infarction. Aliaga J,Bonaventura A, Mezzaroma E, et al. *Molecules* 2021 Jun;26(12):3534
- 108. Targeting the NLRP3 inflammasome in cardiovascular diseases. Toldo S, Mezzaroma E, Buckley LF, et al. *Pharmacol Ther* 2022 Aug; 236:108053
- 109. NLRP3 inflammasome: a novel insight into heart failure. Wang Y, Li Y, Zhang W, et al. *Cardiovasc Transl Res* 2022 Jun
- 110. NLRP3 inflammasome, an immune-inflammatory target in pathogenesis and treatment of cardiovascular diseases. Wang Y, Liu X, Shi H, et al. *Clin Transl Med* 2020 Jan;10(1):91-106
- 111. Apoptosis, pyroptosis, and necrosis: Mechanistic description of dead and dying eukaryoticcells. Fink SL, Cookson BT. *Infect Immunol* 2005; 73)4):1907-16
- 112. Neutrophil extracellular traps in immunity and disease. Papayannopoulos V. *Nat Rev Immunol* 2018 Feb; 18(2):134-47
- 113. Casting a wide NET: An update on uncontrolled NETosis in response to COVID-19 infection. Taylor EB. *Clin Sci (London)* 2022 Jul;136(13):1047-52
- 114. Neutrophil extracellular traps the dark side of neutrophils.Sørensen OE, Borregaard N. *JClin Invest* 2016 May;126(5):1612-20
- 115. Neutrophil extracellular traps: Double-edged swords of innate immunity. Kaplan MJ, RadicM. *J Immunol* 2012 Sep; 189(6):2689-95
- 116. Neutrophil extracellular traps in COVID-19. Zuo Y, Yalavarthi S, Shi H, et al. *JCI Insight* 2020 Jun;5(11):e138999
- 117. Neutrophil extracellular traps contribute to immunothrombosis in COVID-19 acute respiratory distress syndrome.Middleton EA, He XY, Denorme F, et al. *Blood* 2020 Sep;136(10):1169-79
- 118. COVID-19 and neutrophils: The relationship between hyperinflammation and neutrophilextracellular traps.Borges L, Pithon-Curi TC, Curi R, et al. *Mediators Inflamm* 2020 Dec; 2020: 8829674

- 119. Neutrophil extracellular traps and by-products play a key role in COVID-19 pathogenesis, risk factors, and therapy. Thierry AR, Roch B. *J Clin Med* 2929 Sep; 9(9):2942
- 120. Neutrophil extracellular traps participate in cardiovascular diseases: recent experimental and clinical insights. Döring Y, Libby P, Soehnlein O. *Clin Res* 2020 Apr;126(9):1228-41
- 121. SARS-CoV-2-triggered neutrophil extracellular traps mediate COVID-19 pathology. VerasFP, Pontelli MC, Silva CM. 2020 Dec;217(12):e20201129
- 122. The emerging role of neutrophils in the pathogenesis of thrombosis in COVID-19. Iliadi V, Konstantinidou I, Aftzoglou K, et al. *Int J Mol Sci* 2021; 22(10):5368
- 123. A NET-thrombosis axis in COVID-19.Hidalgo A. *Blood* 2020; 136(10):1118-19
- 124. Neutrophil extracellular traps (NETS) in autoimmune diseases: A comprehensive review.Lee KH, Kronbichler A, Park DD, et al. *Autoimmun Rev* 2017; 16(11):1160-73
- 125. The Emerging Role of Neutrophils in the Pathogenesis of Thrombosis in COVID-19.Iliadi V,Konstantinidou I, Aftzoglou K. *Int J Mol Sci* 2021
- 126. Neutrophil extracellular traps contribute to CO-VID-19 Hyperinflammation and humoral autoimmunity. Torres-Ruiz J, Absalón-Aguilar A, Nuñez-Aguirre M, et al. *Cells* 2021; 10(10):2545
- 127. Autoantibodies stabilize neutrophil extracellular traps in COVID-19. Zuo Y, Yalavarthi S, Navaz SA, et al. *JCI Insight* 2021; 6(15):150111
- 128. Neutrophil extracellular traps (NETs) in severe SARS-CoV-2 lung disease. Szturmowicz M, Demkow U. *Int J Mol Sci* 2021; 22(16):6854
- 129. Vasculitis and neutrophile extracellular traps in lungs of Golden Syrian hamsters with SARS-CoV-2. Becker K, Beythien G, de Buhr N, et al. *Front Immunol* 2021; 12: 640842
- 130. Functions of lysosomes.De Duve C, Wattiaux R. *Annu Rev Physiol* 1966; 28:435-492.
- 131. The lysosome turns fifty. De Duve C. *Nat Cell Biol* 2005; 7:847-849



- 132. Historical landmarks of autophagy research. Ohsumi Y. *Cell Res* 2014 Jan;24(1):9-23.
- 133. Autophagy and energy metabolism. Yang J, Zhou R, Ma Z. *Adv Exp Med Biol* 2019;1206:329-357
- 134. Autophagy and the immune system. Kuballa P, Nolte WM, Castoreno AB, Xavier RJ. *AnnuRev Immunol* 2012;30:611-46
- 135. Autophagy in innate and adaptive immunity.Xu Y, Eissa NT. *Proc Am Thorac Soc* 2010;7(1):22-28
- 136. Autophagy in immunity and inflammation.Levine B, Mizushima N, Virgin HW. *Nature* 2011;469(7330):323-35.
- 137. Autophagy and inflammation.Matsuzawa-Ishimoto Y, Hwang S, Cadwell K. *Annu RevImmunol* 2018;36:73-101
- 138. Autophagy: The spotlight for cellular stress response. Ravanan P, Srikumar IF, Talwar P. *Life Sci* 2017;188:53-67
- 139. Alzheimer's Disease: Past, Present, and Future. Bondi MW, Edmonds EC, Salmon DP. *J Int Neuropsychol Soc* 2017 Oct;23(9-10):818-831
- 140. Alzheimer's disease: initial report of the purification and characterization of a novel cerebrovascular amyloid protein.Glenner GG, Wong CW. *BiochemBiophys Res Commun* 1984 May; 120(3):885-90
- 141. Amyloid precursor protein gene mutation in early-onset Alzheimer>s disease. Van Dulin CM, Hendriks L, Cruts M, et al. *Lancet* 1991 Apr; 337(8747):978
- 142. Amyloid deposition as the central event in the aetiology of Alzheimer's disease. Hardy J, Allsop D. *Trends Pharmacol Sci* 1991 Oct; 12(10): 383-88
- 143. Alzheimer's disease and amyloid: culprit or coincidence? Skaper SD. *Int Rev Neurobiol* 2012;102:277-316
- 144. Genetics of b-amyloid precursor protein in Alzheimer's disease. Julia TCW, Goate AM. *ColdSpring Harbor Perspect Med* 2017 Jun; 7(6):a024539
- 145. Amyloid precursor processing and Alzheimer's disease. O'Brien RJ, Wong PC. *Ann RevNeurosci* 2011; 34:185-204

- 146. Alzheimer's disease: as it was in the beginning. Kozlov S, Afonin A, Evsyukov I, et al. *RevNeurosci* 2017 Nov;28(8):825-43
- 147. Genomics of Alzheimer's disease implicates the innate and adaptive immune systems. Li Y, Laws SM, Miles LA et al *J Cell Mol Life Sci* 2021 Dec; 78(23):7397-7426
- 148. Mitochondrial dysfunction in sporadic and genetic Alzheimer's disease. Hauptmann S, KeilU, Scherping I, et al. *Exp Gerontol* 2006 Jul;41(7):668-73
- 149. Autophagy in neurodegenerative diseases: A hunter for aggregates.Park H, Kang JH, Lee S. *J Mol Sci* 2020 May;21(9):3369
- 150. Microglial autophagy defect causes Parkinson disease-like symptoms by acceleratinginflammasome activation in mice. Cheng J, Liao Y, Dong Y, et al. *Autophagy* 2020 Dec;16(12):2193-2205
- 151. The role of autophagy in neurodegenerative disease. Nixon RA. 2013 Aug;19(8):983-97
- 152. Neuronal autophagy and axon degeneration. Rana T, Behl T, Sehgal A, et al. *Cell Mol LifeSci* 2018 Jul;75(13):2389-2406
- 153. Exploring the role of autophagy dysfunction in neurodegenerative disorders.Rana T, BehlT, Sehgal A, et al. *Mol Neurobiol* 2021 Oct;58(10):4886-4905
- 154. Protein Misfolding Diseases.Hartl FU. Annu Rev Biochem. 2017 Jun 20;86:21-26
- 155. Protein folding and misfolding: a paradigm of self-assembly and regulation in complexbiological systems. Vendruscolo M, Zurdo J, MacPhee CE, Dobson CM. *Philos Trans A Math PhysEng Sci.* 2003 Jun 15;361(1807):1205-22.
- 156. Principles of protein folding, misfolding and aggregation. Dobson CM. *Semin Cell Dev Biol* 2004 Feb;15(1):3-16.
- 157. Unraveling the mysteries of protein folding and misfolding. Ecroyd H, Carver JA. *IUBMBLife* 2008 Dec;60(12):769-74
- 158. In vivo aspects of protein folding and quality control.Balchin D, Hayer-Hartl M, Hartl FU.Science 2016 Jul 1;353(6294):aac4354



- 159. The unfolded protein response: mechanisms and therapy of neurodegeneration. SmithHL, Mallucci GR. Brain. 2016 Aug; 139 (Pt 8): 2113-21
- 160. Protein misfolding in the endoplasmic reticulum as a conduit to human disease. WangM, Kaufman RJ.Nature. 2016 Jan 21;529(7586):326-35
- 161. Protein degradation and protection against misfolded or damaged proteins. Goldberg AL. *Nature* 2003 Dec 18;426(6968):895-9.
- 162. Cellular stress responses in protein misfolding diseases. Duennwald ML. *Future Sci OA*. 2015 Sep 1;1(2):FSO42
- 163. Folding proteins in fatal ways. Selkoe DJ. *Nature*. 2003 Dec 18;426(6968):900-4.
- 164. Comparison of four staining methods on the detection of neuritic plaques. WisniewskiHM, Wen GY, Kim KS. *Acta Neuropathol.* 1989;78(1):22-7
- 165. Oligomeric amyloid beta associates with postsynaptic densities and correlates withexcitatory synapse loss near senile plaques.Koffie RM, Meyer-Luehmann M, Hashimoto T, etal. *Proc Natl Acad Sci USA* 2009 Mar; 106(10):4012-17
- 166. The Fourth Phase of Water: Beyond Solid, Liquid, Vapor. Gerald H. PollackEbner& SonsPublishers, 2013
- 167. Water structure and interactions with protein surfaces.Raschke TM. *CurrOpin Struct Biol* 2006 Apr;16(2):152-9
- 168. Water Determines the Structure and Dynamics of Proteins.Bellissent-Funel MC, HassanaliA, Havenith M, Henchman R, Pohl P, Sterpone F, van der Spoel D, Xu Y, Garcia AE. *Chem Rev* 2016 Jul 13;116(13):7673-97
- 169. Water mediation in protein folding and molecular recognition. Levy Y, Onuchic JN. *AnnuRev Biophys-Biomol Struct* 2006;35:389-415.
- 170. Dynamics of hydration water in proteins. Teixeira J. *Gen PhysiolBiophys* 2009;28(2):168-73.
- 171. Sub-terahertz spectroscopy reveals that proteins influence the properties of water atgreater distances than previously detected. Sushko O, Dubrovka R, Donnan RS. *J Chem Phys* 2015 Feb 7;142(5):055101

- 172. Recherches sur la conductivitégalvanique des électrolytes, Svante Arrhenius doctoraldissertation, 1884. Stockholm, Royal publishing house, P. A. Norstedt&Söner
- 173. Aether, fields & energy dynamics in living bodies Part II.Thorp KE, Thorp JA, Walker PR.*G Med Sci* 2021; 2(6): 001-020.
- 174. Species and Specificity: An Interpretation of the History of Immunology. Pauline Mazumdar.publ. Cambridge Univ Press1995: pp. 179-253
- 175. Life at the Cell and Below-Cell Level: The Hidden History of a Fundamental Revolution in Biology. Gilbert Ling. Pacific Press, New York (2001)
- 176. Principles that govern the folding of protein chains. Anfinsen CB. *Science*. 1973 Jul 20;181(4096):223-30.
- 177. How to fold graciously.Levinthal C.Mossbauer Spectroscopy in Biological Systems:Proceedings of a meeting held at Allerton House, Monticello, Illinois: Eds.Debrunner P, Tsibris IC&Münck E, Univ Illinois Press(1969)
- 178. Funnels, pathways, and the energy landscape of protein folding: A synthesis.BryngelsonJD, Onuchic JN, Socci ND, Wolynes PG.Proteins.1995 Mar;21(3):167-95.
- 179. The role of water in amyloid aggregation kinetics. Stephens AD, Kaminski Schierle GS. *CurrOpin Struct Biol* 2019 Oct; 58:115-123
- 180. Role of water in aggregation and amyloid polymorphism. Thirumalai D, Reddy G, Straub JE. *Acc Chem Res* 2012 Jan; 45(1):83-92
- 181. Interaction with surrounding water plays a key role in determining the aggregation propensity of proteins. Chong SH, Ham S. *Angew Chem Int Ed Engl.* 2014 Apr: 53(15):1961-4
- 182. Protein structural and surface water rearrangement constitute major events in the earliestaggregation stages of tau. Pavlova A, Cheng CY, Kinnebrew M, Lew J, Dahlquist FW, Han S. *ProcNatl Sci USA*. 2016 Jan;113(2):E127-36
- 183. Local structure and dynamics of hydration water in intrinsically disordered proteins. RaniP, Biswas P. *J Phys Chem B* 2015 Aug; 119(34): 10858-67



- 184. The amyloid hypothesis on trial.Makin S. *Nature* 2018 (559); July 2018:S4-S7
- 185. Prion diseases. Ironside JW, Ritchie DL, Head MW. *Handb Clin Neurol*. 2017;145:393-403
- 186. Prion Disease. Baldwin KJ, Correll CM. Semin Neurol. 2019 Aug;39(4):428-439
- 187. Infectious and Sporadic Prion Diseases. Richard Knight. *Prog Mol Biol Transl Sci* 2017;150,293-318
- 188. Cellular and molecular mechanisms of prion disease. Sigurdson CJ, Bartz JC, Glatzel M. *Annu Rev Pathol* 2019 Jan;14:497-516
- 189. Prion protein misfolding.Kupfer L, Hinrichs W, Groschup MH. *Curr Mol Med*2009 Sep;9(7):826-35
- 190. COVID-19 outcomes in hospitalized Parkinson's disease patients in two pandemic waves in 2020: A nationwide cross-sectional study from Germany. Scherbaum R, Bartig D, Richter D, et al. *Neurol Res Pract* 2022 Jul; 4:27 https://neurolrespract.biomed-central.com/articles/10.1186/s42466-022-00192-x
- 191. Prevalence and impact of COVID-19 in Parkinson's disease: evidence from a multi-centersurvey in Tuscany region.Del Prete E, Francesconi A, Palermo G. *J Neurol* 2021 Apr;268(4):1179-87
- 192. COVID-19 Vaccine Associated Parkinson's Disease, A Prion Disease Signal in the UK YellowCard Adverse Event Database.Classen JB. *J Med Clin Res & Rev* 2021; 5(7):1-6
- https://scivisionpub.com/pdfs/covid19-vaccine-associated-parkinsons-disease-a-prion-disease-signal-in-the-uk-yellow-card-adverse-event-database-1746.pdf
- 193) COVID-19 and Parkinsonism: A critical appraisal. Cavallieri F, Fioravanti V, Bove F, et al. *Biomolecules* 2022 Jul;12(7):970
- 194. Possible link between SARS-CoV-2 infection and Parkinson's disease: the role of toll-likereceptors. Conte C. *Int J Mol Sci* 2021 Jul;22(13):7135
- 195. Parkinson's disease and the COVID-19 pandemic.Fearon C, Fasano A. *Parkinson's Dis* 2021;11(2):431-44
- 196. The Impact of the COVID-19 Pandemic on Alzheimer's Disease and Other Dementias. Gan J,Liu S,

- Wu H, et al. Front Psychiatry 2021 Jul; 12:703481
- 197. COVID-19 and dementia: Analyses of risk, disparity, and outcomes from electronic healthrecords in the US.Wang QQ, Davis PB, Gurney ME, et al. *Alzheimer's Dement* 2021 Aug;17(8):1297-1306
- 198. The effects of the COVID-19 pandemic on people with dementia. Numbers K, Brodaty H. *Nat Rev Neurol* 2021 Jan; 17:69-70
- 199. A case of probable Parkinson's disease after SARS-CoV-2 infection. Cohen ME, Eichel R, Steiner-Birmanns, et al. *Lancet Neurol* 2020 Oct; 19(10): 804-05
- 200. Creutzfeldt-Jakob disease in a man with COVID-19: SARS-CoV-2-accelerated neurodegeneration? Young MJ, O'Hare M, Matiello M, et al. *Brain BehavImmun* 2020 Oct; 89:601-603
- 201. Creutzfeldt-Jakob disease after COVID-19 vaccination.Kuvandik A, Özcan E, Serin S, et al. *Turk J Intensive Care*. https://cms.galenos.com.tr/Uploads/Article_50671/TYBD-0-0.pdf
- 202. Rapidly Progressive Dementia with Asymmetric Rigidity Following ChAdOx1 nCoV-19 Vaccination. Chakrabarti SS, Tiwari A, Jaiswal S, et al. *Aging Dis*2022 Jun; 13(3):633-36 https://pubmed.ncbi.nlm.nih.gov/35656106/
- 203. Purified and synthetic Alzheimer's amyloid beta (Aβ) prions. Stohr J, Watts JC, MensingerZL, et al. *Proc Natl Acad Sci USA* 2012; 109: 11025–11030.
- 204. Transmission and spreading of tauopathy in transgenic mouse brain. Clavaguera F,Bolmont T, Crowther RA, et al. *Nat Cell Biol* 2009;11: 909–913.
- 205. Amyloidogenesis of SARS-CoV-2 Spike ProteinNyström S, Hammarström P. *J Am Chem Soc* 2022;144(20):8945-50
- 206. Presence of a SARS-COV-2 protein enhances Amyloid Formation of Serum Amyloid. A JanaAK, Greenwood AB, Hansmann UHE. *J Phys Chem B.* 2021 Aug;125(32):9155-67 https://pubmed.ncbi.nlm.nih.gov/34370466/
- 207. Interactions between SARS-CoV-2 N-protein and a-synuclein accelerates amyloidformation. Semerdzhiev SA, Fakhree MAA, Segers-Nolten I, et al. *ACS Chem Neurosci* 2022;13(1):143-50



- 208. SARS-CoV-2 Proteins Interact with Alpha Synuclein and Induce Lewy Body-like Pathology InVitro. Wu Z, Zhang X, Huang Z, Ma K. *Int J Mol Sci* 2022 Mar; 23(6):3394
- 209. Protein Misfolding in Prion and Prion-like diseases: reconsidering a required role forprotein loss of function. Leighton P, Allison WT. *J Alzheimer's Dis.* 2016
- 210. Intercellular spread of protein aggregates in neurodegenerative disease. Davis AA, Leyns C, Holtzman DM. *Annu Rev Cell Biol.* 2018
- 211. Involvement of gut-associated lymphoid tissue of ruminants in the spread of transmissiblespongiform encephalopathies. Press CM, Heggebø R, Espenes A.Adv Drug Deliv Rev. 2004 Apr 19;56(6):885-99.
- 212. Prions and their lethal journey to the brain.Mabbott NA, MacPherson GG.Nat RevMicrobiol. 2006 Mar;4(3):201-11.
- 213. Is alpha-synuclein in the colon a biomarker for premotor Parkinson's disease? Evidencefrom 3 cases. Shannon KM, Keshavarzian A, Dodiya HB, Jakate S, Kordower JH.MovDisord. 2012 May;27(6):716-9
- 214. Alpha-synuclein pathology of the spinal and peripheral autonomic nervous system inneurologically unimpaired elderly subjects. Bloch A, Probst A, Bissig H, Adams H, Tolnay M. Neuropathol Appl Neurobiol. 2006 Jun;32(3):284-95.
- 215. Alpha-Synuclein in Peripheral Tissues in Parkinson's Disease.Ma LY, Liu GL, WangDX, Zhang MM, Kou WY, Feng T. ACS Chem Neurosci. 2019 Feb 20;10(2):812-823
- 216. Do alpha-synuclein aggregates in autonomic plexuses predate Lewy body disorders? A cohort study.Minguez-Castellanos A, Chamorro CE, Escamilla-Sevilla F, Ortega-Moreno A, Rebollo AC, Gomez-Rio M, Concha A, Munoz DG.Neurology. 2007 Jun 5;68(23):2012-8.
- 217. Gastrointestinal manifestations in Parkinson's disease: prevalence and occurrence beforemotor symptoms. Cersosimo MG, Raina GB, Pecci C, Pellene A, Calandra CR, GutiérrezC, Micheli FE, Benarroch EE.J Neurol. 2013 May;260(5):1332-8
- 218. Autonomic involvement in Parkinson's disease:

- Pathology, pathophysiology, clinical features and possible peripheral biomarkers. Cersosimo MG, Benarroch EE. J. Neurol Sci. 2012 Feb 15;313(1-2):57-63
- 219. Caudo-rostral brain spreading of α-synuclein through vagal connections.Ulusoy A, RusconiR, Pérez-Revuelta BI, Musgrove RE, Helwig M, Winzen-Reichert B, Di Monte DA.EMBO Mol Med. 2013 Jul;5(7):1119-27
- 220. Autonomic dysfunction in Parkinson disease and animal models.Metzger JM, Emborg ME.Clin Auton Res.2019 Aug;29(4):397-414
- 221. Parkinson Disease.Homayoun H.Ann Intern Med. 2018 Sep 4;169(5):ITC33-48
- 222. Constipation in Parkinson's Disease.Stocchi F, Torti M.*Int Rev Neurobiol.* 2017;134:811-826
- 223. Autonomic Dysfunction in Parkinson's Disease: Cardiovascular Symptoms, Thermoregulation, and Urogenital Symptoms. Jost WH. Int Rev Neurobiol. 2017;134:771-785
- 224. Olfaction in Parkinson's disease and related disorders. Doty RL. *Neurobiol Dis.* 2012Jun;46(3):527-52
- 225. Olfactory dysfunction as an early biomarker in Parkinson's disease. Fullard ME, Morley JF, Duda JE. *Neurosci Bull.* 2017 Oct;33(5):515-25
- 226. Olfactory dysfunction in neurodegenerative diseases. Fullard ME, Morley JF, Duda JE, et al. *J Curr Allergy Asthma Rep.* 2018 June 15;18(8):42
- 227. Olfactory dysfunction in aging and neurodegenerative diseases. Dan X, Wechter N, Gray S, et al. *Ageing Res Rev* 2021 Sep;70:101416
- 228. Different prion conformers target the olfactory pathway in sporadic Creutzfeldt-Jakobdisease.Zanusso G, Ferrari S, Benedetti D, et al. *Ann NY Acad Sci* 2009 Jul;1170:637-43
- 229. The human olfactory system in two proteinopathies: Alzheimer's and Parkinson's diseases. Ubeda-Bañon I, Saiz-Sanchez D, Flores-Cuadrado A, et al. *TranslaNeurodegen* 2020 Jun 3;9(1):22
- 230. Neuroanatomy and pathology of sporadic Parkinson's disease.Braak H, Del Tredici K. Adv Anat Embryol Cell Biol. 2009;201:1-119.



- 231. Parkinson's disease: the dual hit theory revisited. Hawkes CH, Del Tredici K, Braak H.Ann NY Acad Sci. 2009 Jul;1170:615-22
- 232. A timeline for Parkinson's disease. Hawkes CH, Del Tredici K, Braak H. Parkinsonism Relat Disord. 2010 Feb;16(2):79-84
- 233. Autonomic dysfunction in neurodegenerative dementias. Idiaquez J, Roman GC. *J NeurolSci.* 2011 Jun 15;305(1-2):22-7
- 234. Autonomic failure in neurodegenerative disorders. Kaufmann H, Biaggioni I. *Semin Neurol* 2003 Dec;23(4):351-63.
- 235. Autonomic dysfunction in Parkinson's disease. Micieli G, Tosi P, Marcheselli S, Cavallini A.*Neurol Sci.* 2003 May;24 Suppl 1:S32-4.
- 236. The prevalence of autonomic symptoms in dementia and their association with physicalactivity, activities of daily living and quality of life. Allan L, McKeith I, Ballard C, Kenny RA. *Dement Geriatr Cogn Disord.* 2006;22(3):230-7
- 237. COVID-19: Post-vaccine Smell and Taste Disorders: Report of 6 Cases. Lechien JR, DialloAO, Dachy B, et al. *Ear Nose Throat J.* 2021 Sep 1;1455613211033125.
- 238. Olfactory and gustatory dysfunctions as a clinical presentation of mild-to-moderate formsof the coronavirus disease (COVID-19): A multicenter European study. Lechien JR, Chiesa-Estomba CM, De Siati DR, et al. *Eur Arch Otorhinolaryngol* 2020:277(6):2251-61
- 239. Olfactory and gustatory dysfunctions due to coronavirus disease (COVID-19): a review ofcurrent evidence.Mehraeen E, Behnezhad F, Salehi MA, et al. *Otorhinolaryngol* 2021 Feb;278(2):307-12
- 240. Olfactory and gustatory dysfunctions in patients with laboratory-confirmed COVID-19infection: A change in the trend.Lal P, Chamoli P, Tuli IP, et al. *Indian J Otolaryngol Head NeckSurg* 2021 July 18:1-7
- 241. Post-COVID-19 vaccine parosmia: A case report. Zamzami OS, Kabli AF, Alhothali AS, et al. *Cureus* 2021 Dec 9; 13(12):e20292
- 242. Olfactory and taste disorders in COVID-19: A systematic review.Costa KVTD, CarnaúbaATL,

- Rocha KW, et al. *Braz J Otorhinolaryngol* 2020 Nov-Dec;86(6):781-92
- 243. Olfactory and gustatory dysfunctions in 100 patients hospitalized for COVID-19: Sexdifferences and recovery time in real-life.Meini S, Suardi LR, Busoni M, et al. *Eur Arch Otorhinolaryngol* 2020 Dec;277(12):3519-23
- 244. Smell and taste alterations in COVID-19: A cross-sectional analysis of different cohorts.Paderno A, Schreiber A, Grammatica A, et al. *Int Forum Allergy Rhinol* 2020 Aug;10(8):955-62
- 245. Autonomic dysfunctionin non-critically ill CO-VID-19 patients during the acute phase of disease. Scala I, Bellavia S, Luigetti M, et al. *Neurol Sci* 2022 Aug; 43(8):4635-43
- 246. Alteration of autonomic nervous system is associated with severity and outcomes inpatients with COVID-19. Pan Y, Yu Z, Yuan Y, et al. *Front Physiol* 2021 May; 12:630038
- 247. Dysautonomia and implications for anosmia in long COVID-19 disease. Vallée A. *J Clin Med* 2021 Nov;10(23):5514
- 248. Autonomic dysfunction in patients with COV-ID-19. Erdal Y, Atalar AC, Gunes T, et al. *Acta Neurol Belg.* 2022 Aug; 122(4):885-91
- 249. Coronary Microvascular dysfunction pathophysiology in COVID-19.Yin J, Wang S, Liu Y, etal. *Microcirculation*. 2021 Oct; 28(7):e12718
- 250. Potential autonomic nervous system dysfunction in COVID-19 patients detected by heartrate variability is a sign of SARS-CoV-2 neurotropic features. Mohammadian M, Golchoobian R. *Mol Biol Rep.* 2022 Aug; 49(8):8131-8137
- 251. Autonomic dysfunction in SARS-CoV-2 infection: acute and long-term implications COVID-19. Editor's page series. Becker RC. *J Thromb Thrombolysis*. 2021Oct; 52(3):692-707
- 252. Post-acute sequelae of COVID-19 and cardiovascular autonomic dysfunction: What do weknow?Bisaccia G, Ricci F, Recce V, et al. *J Cardiovasc Dev Dis* 2021 Nov; 8(11):156
- 253. Post-COVID-19 syndrome: epidemiology, diag-



- nostic criteria and pathogenic mechanisms involved. Carod-Artel FJ. *Rev Neurol* 2021 Jun; 72(11):384-96
- 254. Dysautonomia in COVID-19 patients: a narrative review on clinical course, diagnostic and therapeutic strategies. Carmona-Torre F, Mínguez-Olaondo A, López-Bravo A, et al. *Front Neurol* 2022 May; 13:886609
- 255. Postural orthostatic tachycardia syndrome (POTS) and other autonomic disorders after COVID-19 infection: a case series of 20 patients. Blitshteyn S, Whitelaw S. *Immunol Res* 2021Apr; 69(2):205-211
- 256. Biologically Closed Electric Circuits. Björn Nordenström. Nordic Publications 1983
- 257. SARS-CoV-2 spike protein S1 induces fibrin(ogen) resistant to fibrinolysis: implications for microclot formation in COVID-19.Grobbelaar LM, Venter C, Vlok M, et al. *Biosci Rep* 2021 Aug 27;41(8):BSR20210611
- 258. COVID-19: the rollercoaster of fibrin(ogen), D-dimer, Von Willebrand factor, P-selectin and their interactions with endothelial cells, platelets, and erythrocytes. Grobler C, Maphumulo SC, Grobbelaar LM, et al. *Int J Mol Sci* 2020 Jul 21;21(14):5168
- 259. Prevalence of readily detected amyloid blood clots in 'unclotted' type 2 diabetes mellitus and COV-ID-19 plasma: a preliminary report.Pretorius E, Venter C, Laubscher GJ, et al. *Cardiovasc Diabetol* 2020 Nov 17;19(1):193
- 260. Amyloidogenesis of SARS-CoV-2 Spike Protein. Nyström S, Hammarström P. *J Am Chem Soc* 2022;144(20):8945-50
- 261. SARS-CoV-2 spike protein interactions with amyloidogenic proteins. Potential clues toneurodegeneration. Idrees D, Kumar V. *BiochemBiophys Res Commun* 2021;554:94-98
- 262. Neurotoxic amyloidogenic peptides in the proteome of SARS-COV2: potential implications for neurological symptoms in COVID-19. Charnely M, Islam S, Bindra GK, et al. *Nat Commun* 2022; 13(1):3387
- 263. Prion-like Domains in Spike Protein of SARS-CoV-2 Differ across Its Variants and Enable Changes in Affinity to ACE2. Tetz G, Tetz V. *Microorganisms*

- 2022 Jan; 10(2):28
- 264. Prion protein fragment (106-126) induces prothrombotic state by raising platelet intracellular calcium and microparticle release. Mallick RL, Kumari S, Singh N, et al. *Cell Calcium* 2015 Apr; 57(4):300-11
- 265. Prion protein misfolding. Kupfer L, Hinrichs W, Groschup MH. *Curr Mol Med* 2009 Sep; 9(7): 826-35
- 266. The structure of human prions: from biology to structural models-considerations and pitfalls. Acevedo-Morantes CY, Wille H. *Viruses* 2014 Oct;6(10):3875-92
- 267. Prion protein: the molecule of many forms and faces. Kovač V, Šerbec VC. *Int J Mol Sci* 2022 Jan;23(3):1232
- 268. Formation and properties of amyloid fibrils or prion protein. Yamaguchi KI, Kuwata K. *Biophys Rev*2018 Apr;10(2):517-25
- 269. Prions and Prion-like proteins. Fraser PE. *J Biol Chem* 2014 Jul; 289(29):19839-40
- 270. Recent progress in prion and prion-like protein aggregation. Yi CW, Xu WC, Chen J, et al. *Acta BiochimBiophys Sin (Shanghai)* 2013 Jun;45(6):520-26
- 271. Prion protein as a toxic acceptor of Amyloid-b oligomers. Purro SA, Nicoll AJ, Collinge J. *Biol Psychiatry* 2018 Feb; 83(4):358-68
- 272. Preferential recruitment of conformationally distinct amyloid-b oligomers by the intrinsically disordered region of the human prion protein. Madhu P, Mukhopadhyay S. *ACSChem Neurosci.* 2020 Jan; 11(1):86-98
- 273. Binding between prion protein and Aβ oligomers contributes to the pathogenesis of Alzheimer's disease.Kong C, Xie H, Gao Z, et al. *Virol Sin* 2019 Oct;34(5):475-88
- 274. The role of crowded physiological environments in prion and prion-like protein aggregation.Ma Q, Hu JY, Chen J, et al. *Int J Mol Sci* 2013 Oct; 14(11):21339-52
- 275. Macromolecular crowding favors the fibrillization of β 2-microglobulin by accelerating the nucleation step and inhibiting fibril disassembly. Luo XD, Kong FL, Dang HB, et al. *BiochimBiophys Acta*. 2016



- Nov;1864(11):1609-19
- 276. Crowded cell-like environment accelerates the nucleation step of amyloidogenic protein misfolding. Zhou Z, Fan J-B, Zhu H-L, et al. *J Biol Chem* 2009 Oct; 284(44):30148-5
- 277. The contrasting effect of macromolecular crowding on amyloid fibril formation.Ma Q, Fan JB, Zhou Z, et al. *PLoS One* 2012;7(4):e36288
- 278. Protein kinetic stability.Sanchez-Ruiz JM. *Biophys Chem* 2010 May;148(1-3):1-15
- 279. Association of blood viscosity with mortality among patients hospitalized with COVID-19. Choi D, Waksman O, Shaik A, et al. *J Am Coll Cardiol* 2022 Jul; 80(4):316-28
- 280. Blood viscosity of COVID-19 patient: A preliminary report. Joob B, Wiwanitkit V. *Am J Blood Res* 2021 Feb; 11(1):93-95
- 281. Increased blood viscosity and red blood cell ag-

- gregation in patients with COVID-19. NaderE, Nougier C, Boisson C, et al. *Am J Hematol* 2022 Mar; 2022 Mar:283-92
- 282. Biologically Closed Electric Circuits. Björn Nordenström. Nordic Publications 1983
- 283. A central role for amyloid fibrin microclots in long COVID/PASC: origins and therapeutic implications. Kell DB, Laubscher GJ, Pretorius E. *Biochem J* 2022;479(4):537-59
- 284. Persistent clotting protein pathology in Long COVID/Post-Acute sequelae of COVID-19 (PASC) is accompanied by increased levels of antiplasmin.Pretorius E, Vlok M, Venter C, et al. *Cardiovasc Diabetol* 2021 Aug 23; 20(1):172
- 285. Prevalence of symptoms, comorbidities, fibrin amyloid microclots and platelet pathology in individuals with Long COVID/ Post-Acute Sequelae of COVID-19 (PASC). Pretorius E, Venter C,Laubscher GJ, et al. *ScienceOpen.com* 2022 May.